



Serial Review: Heme Oxygenase in Human Disease  
Serial Review Editor: Phyllis A. Dennery

## Heme oxygenase and the cardiovascular–renal system<sup>☆</sup>

Nader G. Abraham<sup>a,b,\*</sup>, Attallah Kappas<sup>b</sup>

<sup>a</sup>New York Medical College, Basic Science Building, Valhalla, NY 10595, USA

<sup>b</sup>The Rockefeller University Hospital, 1230 York Avenue, New York, NY 10021, USA

Received 9 December 2004; revised 4 March 2005; accepted 5 March 2005

Available online 30 March 2005

### Abstract

Heme oxygenase (HO) has been shown to be important for attenuating the overall production of reactive oxygen species (ROS) through its ability to degrade heme and to produce carbon monoxide (CO), biliverdin/bilirubin, and the release of free iron. Excess free heme catalyzes the formation of ROS, which may lead to endothelial cell (EC) dysfunction as seen in numerous pathological conditions including hypertension and diabetes, as well as ischemia/reperfusion injury. The upregulation of HO-1 can be achieved through the use of pharmaceutical agents, such as metalloporphyrins and some HMG-CoA reductase inhibitors. Among other agents, atrial natriuretic peptide and donors of nitric oxide (NO) are important modulators of the heme–HO system, either through induction of HO-1 or the biological activity of its products. Gene therapy and gene transfer, including site- and organ-specific targeted gene transfer, have become powerful tools for studying the potential role of HO-1/HO-2 in the treatment of various cardiovascular diseases as well as diabetes. HO-1 induction by pharmacological agents or gene transfer of human HO-1 into endothelial cells (ECs) *in vitro* increases cell-cycle progression and attenuates Ang II, TNF-, and heme-mediated DNA damage; administration *in vivo* acts to correct blood pressure elevation following Ang II exposure. Moreover, site-specific delivery of HO-1 to renal structures in spontaneously hypertensive rats (SHR), specifically to the medullary thick ascending limb of the loop of Henle (mTALH), has been shown to normalize blood pressure and provide protection to the mTAL against oxidative injury. In other cardiovascular situations, delivery of human HO-1 to hyperglycemic rats significantly lowers superoxide (O<sub>2</sub><sup>-</sup>) levels and prevents EC damage and sloughing of vascular EC into the circulation. In addition, administration of human HO-1 to rats in advance of ischemia/reperfusion injury considerably reduces tissue damage. The ability to upregulate HO-1 through pharmacological means or through the use of gene therapy may offer therapeutic strategies for cardiovascular disease in the future. This review discusses the implications of HO-1 delivery during the early stages of cardiovascular system injury or in early vascular pathology and suggests that pharmacological agents that regulate HO activity or HO-1 gene delivery itself may become powerful tools for preventing the onset or progression of certain cardiovascular pathologies.

© 2005 Elsevier Inc. All rights reserved.

*Abbreviations:* AA, arachidonic acid; Ang II, angiotensin II; ARF, acute renal failure;  $\beta$ -AR,  $\beta$ -adrenergic receptor; BP, blood pressure; cdk, cyclin-dependent kinases; CEC, circulating endothelial cell; cGMP, cyclic guanosine monophosphate; CKI, cyclin kinase inhibitors; CO, carbon monoxide; CoPP, cobalt protoporphyrin; CORM, carbon monoxide-releasing molecule; COX, cyclooxygenase; CPE, chronic pulmonary emphysema; cTAL, cortical thick ascending limb; CYP450, cytochrome P450; EC, endothelial cell; EC-SOD, extracellular superoxide dismutase; EETs, epoxyeicosatrienoic acids; EGF, endothelial growth factor; eNOS, endothelial nitric oxide synthase; GFR, glomerular filtration rate; GSH, glutathione; HDGF, hepatic-derived growth factor; HETE, hydroxyeicosatetraenoic acid; HIF-1 $\alpha$ , hypoxia inducible factor-1-alpha; HO, heme oxygenase; HPH, hypoxic pulmonary hypertension; HPVSR, hypoxic pulmonary vascular structural remodeling; IMCD, inner medullary collecting duct; iNOS, inducible nitric oxide synthase; JGA, juxtaglomerular apparatus; LDL, low-density lipoprotein; LPS, lipopolysaccharide; MSC, mesenchymal stem cells; mTALH, medullary thick ascending limb of the loop of Henle; NOD, nonobese diabetic; NO, nitric oxide; NOS, nitric oxide synthase; O<sub>2</sub><sup>-</sup>, superoxide; PETN, pentaerythrityl tetranitrate; PG, prostaglandin; PKC, protein kinase C; ROS, reactive oxygen species; sGC, soluble guanylate cyclase; SD, Sprague-Dawley; SHR, spontaneously hypertensive rat; SMC, smooth muscle cell; SnMP, tin mesoporphyrin; SnPP, tin protoporphyrin; STZ, streptozotocin; TALH, thick ascending limb of the loop of Henle; TNF, tumor necrosis factor; TX, thromboxane; VEGFR1, vascular endothelial growth factor receptor I; VEGF, vascular endothelial growth factor; VSMC, vascular smooth muscle cell; ZnDPBG, zinc deuteroporphyrin 2,4-bisglycol; ZnPP, zinc protoporphyrin.

<sup>☆</sup> This article is part of a series of reviews on Heme Oxygenase in Human Disease. The full list of papers may be found on the home page of the journal.

\* Corresponding author. New York Medical College, Basic Science Building, Valhalla, NY 10595, USA.

E-mail address: [Nader\\_abraham@nycmc.edu](mailto:Nader_abraham@nycmc.edu) (N.G. Abraham).

## Contents

|   |    |
|---|----|
| Introduction . . . . .  | 2  |
| HO-derived bilirubin and its antioxidant effect . . . . .                                       | 3  |
| HO-derived CO, vasodilation and antiapoptotic effects . . . . .                                 | 4  |
| HO-derived CO, bilirubin, and balloon angioplasty . . . . .                                     | 4  |
| HO-1 promoter polymorphism. . . . .   | 6  |
| Cytoprotective effects of HO-derived CO and bilirubin following myocardial infarction . . . . . | 6  |
| Carbon monoxide-releasing molecules. . . . .  | 6  |
| HO-1/HO-2, renal function and hypertension . . . . .  | 7  |
| HO-arachidonic acid metabolism: Role of CYP-AA metabolism . . . . .                             | 9  |
| HO-arachidonic acid metabolism: Role of cyclooxygenases. . . . .                                | 10 |
| HO and endothelial cell-cycle progression in diabetes. . . . .                                  | 11 |
| HO-1 and endothelial cell dysfunction in diabetes. . . . .                                      | 13 |
| HO and pharmacologic/genetic interventions in cardiovascular disease . . . . .                  | 13 |
| A. Cardiovascular drugs and HO-1 induction . . . . .  | 13 |
| B. Rationale for gene transfer . . . . .  | 14 |
| C. Gene transfer to correct for cardiovascular disorders . . . . .                              | 14 |
| D. Gene transfer and hypertension . . . . .   | 15 |
| E. Gene transfer and diabetes . . . . .   | 15 |
| F. Gene targeting. . . . .  | 18 |
| Concluding remarks . . . . .  | 18 |
| Acknowledgments . . . . .   | 19 |
| References . . . . .  | 19 |

## Introduction

The heme–heme oxygenase (HO) system is a regulator of endothelial cell (EC)<sup>1</sup> integrity and oxidative stress. Heme is the prosthetic group of numerous enzymes and is important to EC function through regulation of the activity of soluble guanylate cyclase (sGC), nitric oxide synthase (NOS), cytochrome P450 (CYP450) monooxygenases, cyclooxygenase (COX), and catalase [1]. HO exists as the constitutive isoenzymes HO-1, HO-2, and HO-3 [2]. HO-3 has no activity and is not expressed in humans (G. Scapagnini, personal communication). HO-2 is a constitutive isoform [3–5]. HO-1 is an inducible isoform that may be induced through the use of various pharmaceutical agents. We have shown that overexpression of HO can be successfully achieved through site- and organ-specific gene delivery by means of adenoviral, retroviral, and liposome-based vectors [6–11].

HO-1 and HO-2 are both viewed as playing a major role in heme breakdown [1,12] and are alike in terms of mechanism of heme oxidation, cofactor, and substrate specificities, and susceptibility to inhibition by synthetic metalloporphyrins, in which the central iron atom is replaced, for example, by zinc, tin, or other elements (reviewd in [1]). Synthetic heme analogues have been used in animals and in humans to suppress or induce HO activity and to inhibit development of severe hyperbilirubinemia in newborns [13–15]. The recognition that HO-1 is strongly induced by oxidant stress and its substrate, heme, in conjunction with the robust ability of

HO-1 to protect against oxidative insult [7,16–19] has led to examination of the antioxidant nature of HO-1 and HO-2 activities [1,7,18,20]. Antioxidant effects arise from the capacity of HO-1 to degrade the heme moiety from destabilized heme proteins [21] and from biliverdin and bilirubin, products of HO with potent antioxidant properties (Fig. 1) [22,23].

Carbon monoxide (CO), an HO product as well, is not an antioxidant [24], but can cause induction of antioxidant genes; it also decreases superoxide (O<sub>2</sub><sup>-</sup>) levels [25,26], increases GSH levels [27], and has an anti-apoptotic effect [28]. Further, CO is a vasodilator, which has been shown to play an important role in the regulation of basal and constrictor-induced vascular tone in blood vessels [29–34]. Upregulation of bilirubin and CO, through the induction of HO-1, has shown promising results in protection against oxidative stress and injury whereas the absence of HO-1, for example, in mice, results in accelerated atherosclerotic lesion formation and vein-graft disease as well as elevated blood pressure, cardiac hypertrophy, and acute renal failure. In addition, the occurrence of organ damage and mortality are more frequent in HO-1 null mice [35]. Induction of HO-1 also prevents cell death, which is attributed to its augmentation of ion efflux and exportation of iron-binding protein [36]. The protective actions of HO-1 or CO are not confined to overtly oxidant processes, but rather extend widely to such pathologic processes. Connors et al. [37] were the first to demonstrate that induction of HO-1 has an anti-inflammatory effect. This finding was substantiated by studies from other laboratories [38,39]. Further-

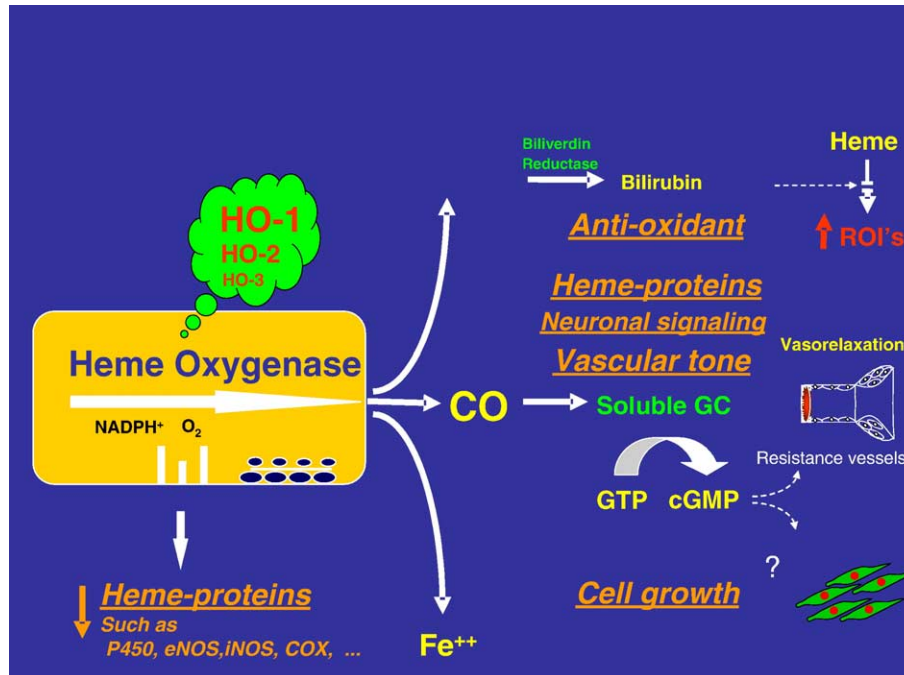


Fig. 1. The heme–HO system: proposed mechanism for HO function in cellular protection following oxidative injury.

more, HO-1 induction has been shown to be cytoprotective in atherosclerosis [40–42,40] sepsis [24], diabetes [26,43], lung injury [44], occlusive vascular disease, and ischemia [45–47].

### HO-derived bilirubin and its antioxidant effect

HO-1 catalyzes the rate-limiting step in heme degradation to biliverdin. Biliverdin is, in turn, converted into bilirubin by the cytosolic enzyme biliverdin reductase at the expense of NADPH. Unconjugated bilirubin efficiently scavenges singlet oxygen and serves as a reducing agent for certain peroxidases, including horseradish peroxidase and prostaglandin H (PGH) synthase, in the presence of hydrogen peroxide or organic hydroperoxides. Biliverdin and bilirubin are reducing species and hence potential antioxidants.

An adult human produces about 300 mg of bilirubin per day [48,49]. Some 80 to 85% of the bilirubin produced in vivo is derived by heme catabolism of hemoglobin released by aging or damaged erythrocytes [48,49]. The biologic actions of bilirubin may be particularly relevant to prevention of the oxidant-mediated vasoconstrictive actions of tumor necrosis factor (TNF) and angiotensin II (Ang II) [50,51]. Bilirubin, in low concentrations, scavenges reactive oxygen species (ROS) in vitro, reduces oxidant-induced cellular injury, and attenuates oxidant stress in vivo [22,52–61]. Mazza et al. [62] have provided evidence of the antioxidant and cytoprotective effects of bilirubin in attenuating Ang II-mediated EC DNA damage and cell death. Morita et al. [63] showed that Ang II significantly stimulates O<sub>2</sub><sup>-</sup> formation in monocytes, and that exogenously applied

bilirubin suppresses not only O<sub>2</sub><sup>-</sup> formation but also Ang II-enhanced chemotactic activity in monocytes.

Bilirubin has been shown to inhibit NADPH oxidase [64] and protein kinase (PKC) activity [65]; both have been shown to mediate Ang II-induced vascular injury [66,67]. Recently, biliverdin and bilirubin have been shown to preserve EC integrity [68] and prevent EC death and sloughing, to enhance vascular reactivity in diabetic rats [26,69], and to prevent restenosis [70,71]. Bilirubin is also implicated in reducing oxidative stress in experimental diabetes, in part, by increasing the bioavailability of nitric oxide (NO) needed for EC integrity. Bilirubin-mediated inhibition of PKC and NADPH oxidase may be one mechanism by which HO-1 attenuates the diabetes-mediated generation of oxidants and the uncoupling of EC nitric oxide synthase (eNOS). Glucose enhances EC O<sub>2</sub><sup>-</sup> production, leading to increased vascular formation of the NO/O<sub>2</sub><sup>-</sup> reaction product, peroxynitrite. Peroxynitrite oxidizes the active NOS cofactor, tetrahydrobiopterin, to cofactor inactive molecules, such as dihydrobiopterin. This uncouples the enzyme, which then preferentially increases O<sub>2</sub><sup>-</sup> production over NO production [72]. Functional eNOS expression, rather than dysfunctional uncoupled eNOS (Fig. 2), may be increased by HO-1 gene expression. Peroxynitrite stimulates HO-1 in nondiabetic pathological conditions [73] but in diabetes [25,26], due to the glucose-suppressive effect on HO-1 gene expression [74]. Thus, the HO-1 gene expression-mediated increase in extracellular superoxide dismutase (EC-SOD) may protect eNOS from uncoupling, which in turn attenuates the diabetes-mediated generation of oxidants [75,76].

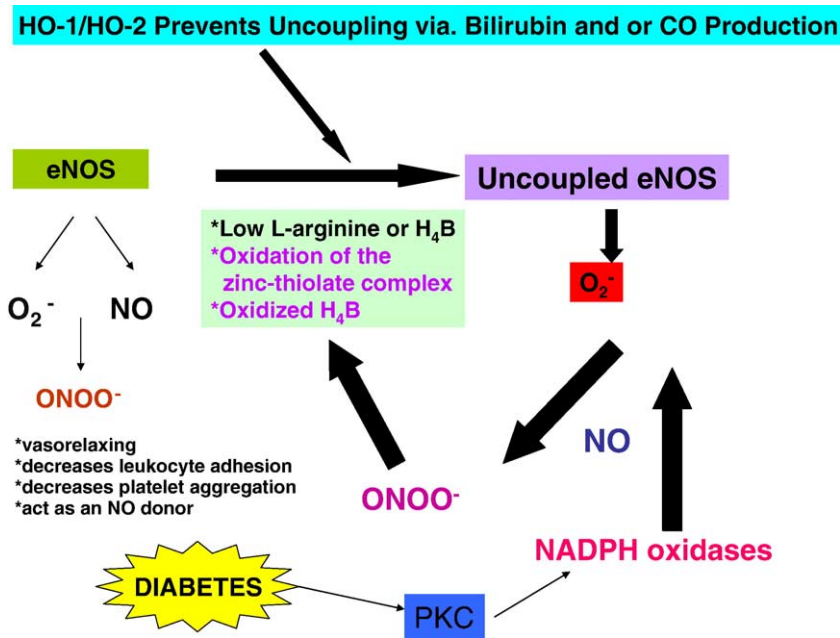


Fig. 2. Schematic representation of the possible interaction of HO/CO in the prevention of eNOS uncoupling.

HO-1-derived bilirubin has also been shown to display cytoprotective properties in the cardiovascular system [61,77]. Numerous reports indicate that a higher serum bilirubin level is associated with a decrease in the risk for coronary artery disease in humans. Free and albumin-bound bilirubin has been shown to inhibit oxidation of low-density lipoprotein (LDL) [54]. In renal vessels from rats pretreated with biliverdin, the lipopolysaccharide (LPS)-induced expression of P- and E-selectin was shown to be significantly attenuated, confirming that biliverdin or its metabolite, bilirubin, might account for the anti-inflammatory properties of HO [78]. Additionally, Wang et al. [79], employing a rat model of endotoxemia, examined the hypothesis that bilirubin is a key mediator of HO-1 cytoprotection. Bilirubin treatment improved survival of cells and attenuated liver injury in response to LPS infusion. Serum levels of NO and  $TNF\alpha$ , key mediators of endotoxemia, and hepatic inducible nitric oxide synthase (iNOS) expression were significantly lower in bilirubin-treated rodents versus control animals. In addition, bilirubin treatment induced a threefold increase in LPS-mediated prostaglandin (PG) synthesis in the absence of significant changes in COX expression or activity, suggesting that bilirubin enhances substrate availability for eicosanoid synthesis.

#### HO-derived CO, vasodilation and antiapoptotic effects

Almost all CO produced in vivo comes from the degradation of heme by HO. Before CO was found to have an antihypertensive effect, Sacerdoti et al. [80] discovered that induction of HO-1 served to attenuate hypertension in

spontaneously hypertensive rats (SHR). This finding provided a potentially crucial lead in defining the potential therapeutic benefits of regulating HO activity in the treatment of hypertension. Later, it was discovered that CO and NO have similar properties [81–83]; both behave as messenger and signaling molecules. Both CO and NO are capable of inducing the relaxation of blood vessels through vasodilation and inhibiting the proliferation of vascular smooth muscle cells (VSMC) [84]. Like NO, HO-derived CO influences the sGC and cyclic guanosine monophosphate (cGMP) pathways, which serve to regulate both blood pressure and vascular contractility [85]. sGC acts to increase cGMP, which in turn serves as a vasodilator to lower blood pressure levels. It has been demonstrated that by upregulating the HO system in young (8-week-old) SHR, coincidentally, sGC and cGMP levels rise, which leads to a significant reduction in blood pressure [85,86]. On the other hand, by using an inhibitor of HO-1 activity, the blood pressure of rats undergoing HO-1 inhibition significantly increases [87]. Moreover, CO has been shown to inhibit platelet aggregation and to stimulate angiogenesis [88]. Interestingly, a study done by Ushiyama et al. [89] indicates that CO regulates blood pressure cooperatively with NO in hypertensive rats and by impairing the function and production of NO, HO gene expression is increased and the effect of CO in the regulation of blood pressure is, in turn, augmented.

#### HO-derived CO, bilirubin, and balloon angioplasty

Apart from being a vasodilator, CO has also been shown to protect against ischemic tissue damage through anti-

apoptotic mechanisms [46]. By inducing HO-1 during postischemic myocardial dysfunction, myocardial function is improved after total ischemia and reperfusion [90,91]. Kukoba et al. [92] injected hemin into rats before inducing ischemia in order to upregulate HO-1. They demonstrated that, during ischemia and reperfusion, left ventricular pressure decreased while end diastolic pressure, coronary perfusion pressure, and coronary resistance increased. The products of HO action are protective in rodent models of ischemia-reperfusion injury, allograft and xenograft survival, intimal hyperplasia following balloon injury, or chronic graft rejection [91]. HO-1 expression is increased in human atherosclerotic lesions [42,93], as well as in vascular EC and smooth muscle cells (SMC) exposed to oxidized LDL [41].

Moreover, Togane et al. [90] have reported that CO generated through HO inhibits mitogen-induced proliferation of VSMCs, which in turn inhibits neointimal formation. According to Tulis et al. [94], the pharmacological induction of HO-1 also attenuates neointima formation after balloon angioplasty-induced injury. Furthermore, they showed that HO-1 gene transfer attenuated the remodeling response to vascular injury by stimulating medial wall SMC apoptosis and inhibiting medial wall DNA replication [94]. Others have shown that adenovirus-mediated HO-1 overexpression inhibits the growth of SMCs in vitro and in vivo, as well as the upregulation of p21<sup>Cip1</sup> [95], suggesting the importance of HO-1 in vascular wall remodeling. Overexpression of human HO-1 in rabbit and rat ECs renders the cells resistant to hemoglobin toxicity and highlights not only the important metabolic and cytoprotective roles of the HO-1 gene [7,17], but also its importance in cell-cycle progression [7,19,96].

Discovery of the differential effects of HO-derived products on ECs and SMCs by Li Volti et al. [96] opens an avenue toward understanding the basic and clinical problems of neointimal formation. Three key findings substantiate this development. The key finding was that induction of HO activity in both ECs and SMCs [50,96,97], as well as other mammalian cells [98,99] regulates cell-cycle progression. The second was that, in SMCs, upregulation of HO-1 gene expression is associated with a decrease in cell-cycle progression. In ECs, unlike SMCs, overexpression of HO-1 increases cell-cycle progression and DNA distribution in the S and G<sub>2</sub>/M phases [25,96]. The third key finding was that inhibition of HO activity with SnMP resulted in the enhancement of cell-cycle progression in SMCs. In contrast, cell-cycle progression was reduced by SnMP in ECs. The use of vinblastine demonstrated that it is possible to distinguish cells with DNA content at different phases of the cell cycle, i.e., G<sub>2</sub>/M cells of low DNA ploidy [100]. The addition of vinblastine to EC and SMC cultures caused cell arrest in mitosis. The percentage of cells entering mitosis thus increases with time, and the rate of the increase reflects the kinetics of cell-cycle progression [101]. HO-1 inhibition increased DNA content in vinblastine-treated SMCs in the G<sub>2</sub>/M phase, suggesting an increased rate of cell proliferation [25,96]. In contrast, in ECs, HO-1 inhibition decreased DNA content in the G<sub>2</sub>/M phase, thus providing evidence of its activating effect on cell-cycle progression [25,102].

In a similar study, McClung et al. [70] also concluded that HO-1 has a differential effect on cell-cycle proliferation in ECs and VSMCs. In their study, inhibition of HO-1 led to VSMC proliferation, thus, causing stenosis in the lumen (Fig. 3B) while induction of HO-1 stimulated neointima

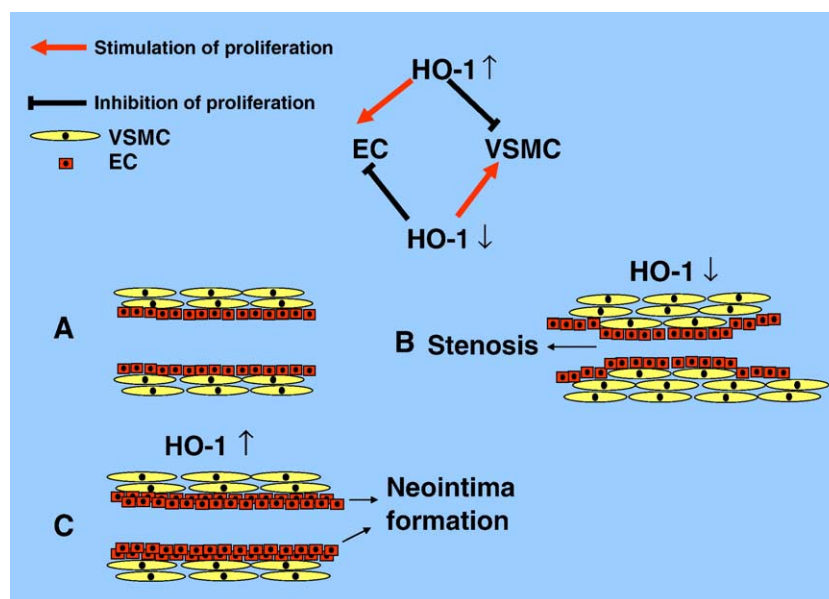


Fig. 3. The possible role of HO/CO in stenosis and neointima formation: a decrease in HO-1 produces vascular smooth muscle cell proliferation and causes stenosis (B). By increasing HO-1, vascular smooth muscle cell proliferation is inhibited and prevents neointima formation (C). This in turn produces healthy tissue (A).

formation (Fig. 3C), compared to control (Fig. 3A). Kong et al. have more recently shown that transplantation of circulating endothelial progenitor cells overexpressing eNOS and HO-1 to balloon-induced injured vessels served to enhance the vasculoprotective properties of the reconstituted endothelium, which in turn led to inhibition of neointimal hyperplasia [70,103]. These findings are of particular relevance since the differential role of HO-1 could be a key factor in vascular wall remodeling in different clinical situations, such as occurs in hypertension or VSMC hyperplasia, or in the recovery of the vascular wall after mechanical injury, such as restenosis.

### HO-1 promoter polymorphism

A series of studies have examined HO-1 polymorphism in human disease. Yamada et al. [93] examined the polymorphism of the HO-1 gene in patients with chronic pulmonary emphysema (CPE). Specifically they examined a (GT)*n* dinucleotide repeat in the 5'-flanking region of the human HO-1 gene and reported that the larger the size of the (GT)*n* repeat in the HO-1 gene promoter the greater the chance of reducing HO-1 inducibility by reactive species in cigarette smoke, resulting in the development of emphysema. Kanai et al. [104] examined the polymorphic (GT)*n* repeat in German and Japanese newborns in an attempt to explain the higher incidence and severity of hyperbilirubinemia in the latter population. A significant difference in the allele frequency of the (GT)*n* repeats between the two populations was found, but they found no relationship between those polymorphisms and neonatal hyperbilirubinemia. Ono et al. [105] reported that the AA genotype of HO-1 is associated with an increased incidence of hypertension in Japanese women. They speculated that CO can attenuate NO-induced vasodilation; thus, a high expression of HO-1 may result in hypertension in women. Kanai et al. [106] found no association of HO-1 polymorphism to susceptibility to Kawasaki disease in Japanese children. Chang et al. [107] examined polymorphism in the HO-1 promoter in relation to the risk of oral squamous cell carcinoma (OSCC) in Asian male areca chewers. They examined the allelotypic frequency of (GT)*n* repeats in 83 controls, 147 OSCC patients, and 71 oral submucous fibrosis patients. Longer (GT)*n* repeat alleles in the HO-1 promoter were associated with the risk of areca-related OSCC, while a shorter (GT)*n* allele may have protective effects, thus confirming the observation of Yanada et al. that the longer the (GT)*n* repeat, the greater the susceptibility to disease. The results above indicate that, in certain disease conditions, the ability of an individual to upregulate HO-1 may be an important protective factor.

The recent elegant work by Funk et al. investigating the association of HO-1 repeat polymorphism and gene expression showed that short GT repeats were associated with upregulation of HO-1 and therefore influenced

susceptibility to ischemic injury [108]. Short repeats (<25 GT repeat) in the HO-1 gene promoter confer reduced risk for a cerebrovascular event in individuals with normal plasma lipid levels [108]. This may explain controversial findings in differing populations. Further, this may also explain the different results of some investigators in different strains of animals. In support of these findings, the work of Schillinger et al. showed that the HO-1 promoter genotype that controls the degree of HO-1 upregulation in responses to stress stimuli is associated with the postintervention inflammatory response and restenosis risk after balloon angioplasty [109].

### Cytoprotective effects of HO-derived CO and bilirubin following myocardial infarction

The induction of HO-1 may also have therapeutic benefits during chronic heart failure. Gu et al. [110] recently demonstrated that upregulation of HO-1 during heart failure serves to mitigate pathologic left ventricular remodeling and reduce myocardial hypertrophy, oxidative stress, and inflammatory activation. Upregulating HO-1 also has the potential of attenuating cardiac hypertrophy in genetically hypertensive rats [111]. More recently, studies by Ooi et al. [112] in mice suggest that HO-1 confers significant antioxidant and cytoprotective effects in the heart. They showed that HO-1 is induced by  $\beta$ -adrenergic receptor ( $\beta$ -AR) stimulation and may protect against  $\beta$ -AR-mediated apoptosis. Further, it has been demonstrated that induction of HO-1 increases adult cardiomyocyte tolerance to ischemia after *in vivo* transplantation [113]. Autologous atrial cardiomyocytes are a readily available cell source for infarct repair; however, they readily undergo apoptosis, precluding their use as cellular repair grafts. This study demonstrates that preconditioning with HO-1 acts to retain functional viability *in vivo* in adult cardiomyocyte cellular grafts after implantation. This, in turn, may prove effective in repairing infarcted myocardium. In addition, Tang et al. [114] have performed similar studies with mesenchymal stem cells (MSC). They demonstrated that hypoxia-regulated HO-1 vector modification enhances the tolerance of engrafted MSCs to hypoxia-reoxygenation injury *in vitro* and improves their viability in an ischemic milieu. These findings may also make cell therapy more effective in infarct repair.

### Carbon monoxide-releasing molecules

The discovery, by Motterlini et al. [115], that transitional metal carbonyls have the ability to bind and release CO has furthered the understanding of CO in vascular biology. Such pharmacological inducers have been termed carbon monoxide-releasing molecules (CORMs) and have contributed greatly to our understanding of the role each HO product

might play in vascular cytoprotection [115,116]. Water-soluble CORM-3 (Ru(CO)<sub>3</sub>Cl(glycinate)) has been recently developed and has been used to confirm the vasodilatory and cardiovascular benefits of CO. CORM-3 (Fig. 4) has the capability of generating aortic vasodilation *ex vivo* and reduces blood pressure *in vivo* [115–118]. Guo et al. showed that the administration of CORM-3 could reduce infarct size *in vivo* when given in a clinically relevant manner at the time of reperfusion [46]. The results indicate that infarct size was dramatically smaller in treated mice. This experiment shows that CORM-3 can be used not only to bypass HO-1 induction but also to provide the beneficial effect needed as evidenced by a decrease in injury during myocardial ischemia–reperfusion [46]. Although the cardioprotective effect of CORMs occurs through the release of CO and may also occur through the induction of HO, it is by means of HO that the degradation of heme releases CO and produces favorable cardiovascular benefits. In a study conducted by Stanford et al. [84], pulmonary artery SMCs showed an elevation of HO-1 in the presence of oxidants. Through the use of the CORM tricarbonyldichlororuthenium (II) dimer, the proliferation of pulmonary artery SMCs was shown to profoundly decrease. It is apparent that the upregulation of CO through HO-1 has a profound opposing effect to hypertension. Rodella et al. (personal communication). have shown that CORM-3 renders ECs resistant to oxidative stressors even when HO-1/HO-2 are inhibited with SnMP.

CO produced from heme metabolism in blood vessels is reported to elicit relaxation [32,119] through the elevation of cGMP levels [120]. Ndisang et al. [121] studied the administration of hemin, an inducer of HO, and its effect on the pulmonary artery in young SHR and demonstrated that hemin decreases blood pressure (BP) by inducing the HO/CO-sGC/cGMP system. Impairment of this system in the pulmonary artery may be indicative of the pathogenesis and development of hypertension. Sacerdoti et al. [80] and Levere et al. [122] have shown that upregulating the HO/CO system lowers BP in young SHR but not in adults. In young SHR, blood pressure rises and continues to increase with aging whereas adult SHR have an established hypertension

[80,121]. It was hypothesized that if the HO/CO system is defective in young SHR, then HO-1 inducers could enhance the activity of this system.

In assessing the effect of CO-releasing molecules, such as CORM-3, it is essential to use a pharmacological dose. Motterlini's group has shown that a physiological dose of CO has a powerful vasodilator effect and attenuates the phenylephrine-contracting effect [118,123,124]. However, nonphysiological levels of CO-releasing molecules will result in less relaxation or contraction [R. Motterlini, personal communication]. This is essentially similar to the effect of acute and chronic HO-1 inducers on cell survival and on heme-dependent enzymes [125,126]. For example, moderate induction of HO-1 increases cGMP, but extreme levels of HO-1 lead to a decrease in cGMP [125], which has been attributed to the stripping of heme from the sGC protein. This is analogous to the roles of COX-2 and NOS; in cell function, for example, moderate levels of COX-2 are beneficial while an extreme elevation in COX-2 can have harmful effects. A similar situation exists with inhibitors of HO. At low levels, these inhibitors can decrease CO and bilirubin; however, at a higher concentration, they can inhibit other heme proteins, including CYP450, NOS, or COX [127].

### HO-1/HO-2, renal function and hypertension

HO is present in the kidney as the constitutive isoform, HO-2, and as the inducible isoform, HO-1. The discovery that atrial natriuretic peptide is an inducer of HO-1 in renal and endothelial cells [128] opens a new direction in the immunoprotection of renal function during the use of cyclosporin [129]. Cyclosporin is an immunomodulator and provides cytoprotection during organ transplant; however, due to the cyclosporine-suppressive effect on HO-1, its use became limited. Rifampin is another immunomodulator and its beneficial effect has been [130] attributed to its ability to enhance HO-1 gene expression. Atrial peptide-mediated upregulation of HO-1 [128,129,131] has been shown to ameliorate the effect of cyclosporine on renal function, presumably via a cGMP-dependent mechanism [131].

Induction of HO-1, by retroviral delivery of the human HO-1 gene into human microvessel endothelial cells, has been shown to attenuate TNF $\alpha$ -mediated cell death [50,51]. HO-1 gene upregulation has also been shown to attenuate Ang II-mediated DNA damage in endothelial and kidney cells [27,62]. CO, in addition to activating the cGMP pathway and eliciting vasodilation [132], has also been shown to function as an anti-inflammatory and cytoprotective molecule in renal tissue. Further, CO has also been shown to participate in the regulation of ion channels, including the apical 70-pS K-channel activity of the thick ascending limb of the loop of Henle (TALH) [133]. It is now believed that CO-induced NO may have a differential effect

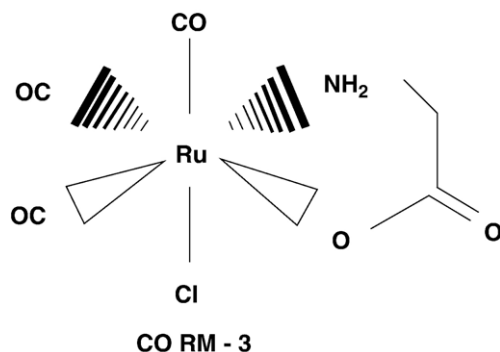


Fig. 4. Schematic representation of the structure of CORM-3 [115].

on cGMP due to the low affinity of sGC for CO; however, it may cause the robust production of cGMP [134].

Abnormalities in kidney function are responsible, in part, for the elevation of blood pressure in SHR. Blood pressure elevation in SHR has been shown to decrease through renal transplantation from normotensive donors [135]. Renal abnormalities leading to hypertension consist of reduced excretion of sodium and water, decreased renal blood flow, and decreased glomerular filtration. Such abnormalities can be corrected by inducing HO-1 activity. HO-1 induction has also been shown to exert a protective effect on renal function in animal models of rhabdomyolysis [136], cisplatin nephrotoxicity [98], and nephrotoxic nephritis [137]. Further, the products of HO provide a protective role in acute renal failure (ARF) and hypertension. Arregui et al. [138] have demonstrated that HO-1-derived CO increased blood carboxyhemoglobin levels, renal blood flow, glomerular filtration, and urinary cGMP excretion. In a similar study, it was verified that the renal vascular bed of hypertensive animals exhibits a greater propensity to upregulate the HO system, which may serve to lower blood pressure in hypertension [139]. In another study [140], it was established that heme-induced renal vasodilation, which increases renal blood flow, is a COX-dependent response whereas heme-induced diuresis and natriuresis are HO-dependent responses, involving inhibition of tubular reabsorption of water and sodium.

A well-known source of oxidative stress in the kidney is the inflammatory cytokine, Ang II. Ang II is systemically or locally elevated in many forms of hypertension [141] and is

associated with increased vascular  $O_2^-$  production [142] which contributes to renal injury [143,144]. Moreover, increased  $O_2^-$  has been shown to contribute to the vascular and renal effects of Ang II [145]. Ang II induces renal oxidant stress and HO activity, suggesting that upregulation of HO-1 in renal proximal tubules is essential in the amelioration of oxidant-mediated injury [143]. In addition, HO-1 is reported to counteract aldosterone-elicited arterial injury through the inhibition of oxidative stress as well as inflammatory reactions [112]. Thus, it is apparent that HO-1 and its products play a critical protective role against cardiovascular injury. Fig. 5 illustrates the roles of bilirubin and CO in preventing DNA degradation, decreasing p21 and p27 levels, and enhancing cell-cycle progression [50,97].

In a rat model of radiation-induced nephropathy, elevated glomerular HO-1 expression was prevented by treatment with AT<sub>1</sub>-receptor antagonists, which block the upregulation of HO-1 expression. This suggests that Ang II may be a mediator of HO-1 induction [146]. Furthermore, Haugen et al. [143] demonstrated elevated levels of HO-1 in the renal proximal tubule of rats treated with Ang II, which was associated with increased HO activity. These data demonstrate that Ang II directly induces HO-1 in renal proximal tubular epithelial cells in vitro. In another study, Ang II infusion decreased glomerular filtration rate (GFR) and increased proteinuria, which led to hypertension. HO-1 upregulation naturally followed and provided a cytoprotective effect [144]. Once again, the evidence suggests that HO-1 has a protective effect against the development of hypertension.

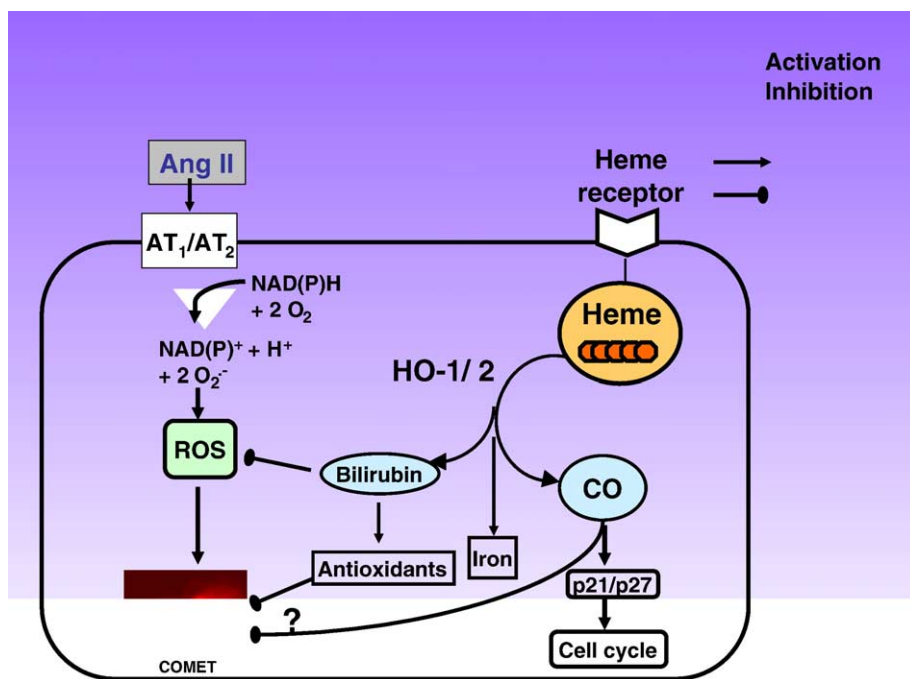


Fig. 5. Schematic representation of the hypothesis that overexpression of HO-1 attenuates the cytostatic effects of Ang II and DNA degradation as measured by COMET assay. CO inhibits p21 and p27 and increases cell-cycle progression while bilirubin inhibits ROS-mediated DNA degradation.



In contrast, Ishizaka and Griendling have shown that treatment of rat VSMCs with Ang II decreased HO-1 mRNA levels and this decrease was blocked by losartan, a selective AT<sub>1</sub>-receptor antagonist [147]. It is conceivable then that Ang II-mediated upregulation of HO-1 subserves mechanisms that counteract the action of Ang II. The recent findings that HO-1 gene transfer ameliorated oxidative tissue injury [9,28] and that oxidant-induced cellular injury was increased in HO-1 knockout mice [148] and in human HO-1 deficiency [149] provide further evidence that HO-1 acts favorably against oxidative stress. Moreover, Ang II, delivered by an osmotic minipump, provoked systemic hypertension, which was associated with an elevation of vascular and renal HO-1 [143,144,150]. Importantly, in animal models, Ang II-induced O<sub>2</sub><sup>-</sup> production and elevated BP were prevented by preadministration of other antioxidant genes, such as SOD [151] or the SOD mimetic tempol [152].

Yang et al. [10] proposed that overexpression of HO-1, in rats transduced with retroviruses containing the human HO-1 gene, would significantly attenuate pressor responsiveness to Ang II and ameliorate Ang II-induced hypertension in rats chronically infused with Ang II (Fig. 6). The postulated mechanisms by which overexpression of HO-1 improves vascular function and ameliorates both genetic and experimental (Ang II-induced) forms of hypertension include the following: the ability of CO to inhibit constrictor responsiveness to myogenic stimuli and attenuate the sensitivity of arterial vessels to vasoconstrictors [32,132]; the capacity of biliverdin and bilirubin, as antioxidants, which may down-regulate the activity of redox mechanisms involved in the vascular actions of Ang II and in the development of hypertension [153]; the potential that HO-1 overexpression minimizes oxidative stress as a result of lowering cellular heme; and the ability of HO activity to affect the expression of COX and CYP450 [154,155]. The production of constrictor eicosanoids, including endoperoxides and thromboxanes as well as 20 HETE [156–159], would be regulated by the levels of HO-1 [80,155,160–162]. The fact that Ang II administration rapidly increases HO-1 expression and HO activity in several tissues [67,144,150] further suggests that

the heme–HO-1 system serves as a control mechanism to the pressor activity of Ang II.

### HO-arachidonic acid metabolism: Role of CYP-AA metabolism

The metabolites of arachidonic acid (AA) metabolism by CYP450-dependent enzymes have been associated with the development and maintenance of hypertension (Fig. 7). When compared to control groups, CYP content and activity are increased in the kidneys of SHR [80,163]. CYP enzymes catalyze the metabolism of AA to four epoxyeicosatrienoic acids (EETs),  $\omega/\omega-1$  alcohols (20-HETE and 19-HETE), and six regioisomeric *cis-trans*-conjugated monohydroxyeicosatetraenoic acids (HETEs) [164,165]. Some of these metabolites (e.g., 5,6-EET and 20-HETE) can be processed further by COX to products having biological activities [166,167]. Metabolites of AA, via the CYP450 pathway, are endowed with biological activities most relevant to the vascular and renal mechanisms of blood pressure regulation. 20-HETE acts as a vasoconstrictor whereas 19-HETE functions to increase sodium retention by acting as a potent sodium-potassium-ATPase stimulator. Furthermore, 20-HETE can stimulate contraction of VSMC [168], inhibit Na<sup>+</sup>-K<sup>+</sup>-ATPase [169], and reduce the activity of potassium channels in arterial smooth muscle and renal tubular cells [170–172]. 20-HETE also affects the movement of ions, constricts blood vessels, participates in tubuloglomerular feedback, and acts as mitogen [173,174], effects that are prohypertensive. Together, both monooxygenases serve to increase blood pressure. In 1990, studies from this laboratory described that hypertension could be attenuated in SHR through administration of heme arginate, a potent inducer of HO-1 [122]. As CYP450 is a heme-containing protein, CYP450 levels are regulated in part by the availability of heme. These experimental data indicate that the increase in HO activity is associated with a parallel decrease in CYP450 content and in the activity of CYP450  $\omega/\omega-1$  arachidonate hydroxylases in SHR kidneys.

HO-derived CO has been reported to inhibit the activity of CYP450 [175] and the generation of vasoconstrictive substances, such as 20-HETE, thus ameliorating the development of hypertension. Sacerdoti et al. [80] used SnCl<sub>2</sub>, a specific inducer of renal HO, to demonstrate that increased HO activity, which resulted in depletion of the CYP-AA metabolites, 20-HETE and 19-HETE, was associated with reduction in BP. In addition, Sabaawy et al. [176] have shown that a single intracardiac injection of retroviral vector containing the human HO-1 gene attenuated the development of hypertension in 5-day-old SHR. Others have shown that administration of heme arginate caused a rapid decrease in BP in young SHR [122,162,177] and that pretreatment with ZnDPBG, a potent HO inhibitor [178], greatly attenuated the antihypertensive response to heme arginate. Thus, it appears

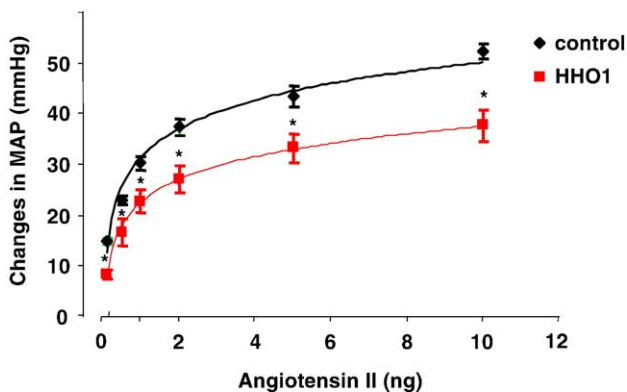


Fig. 6. Angiotensin II-induced blood pressure increase is attenuated in rats expressing the human HO-1 gene using retrovirus gene transfer [10].

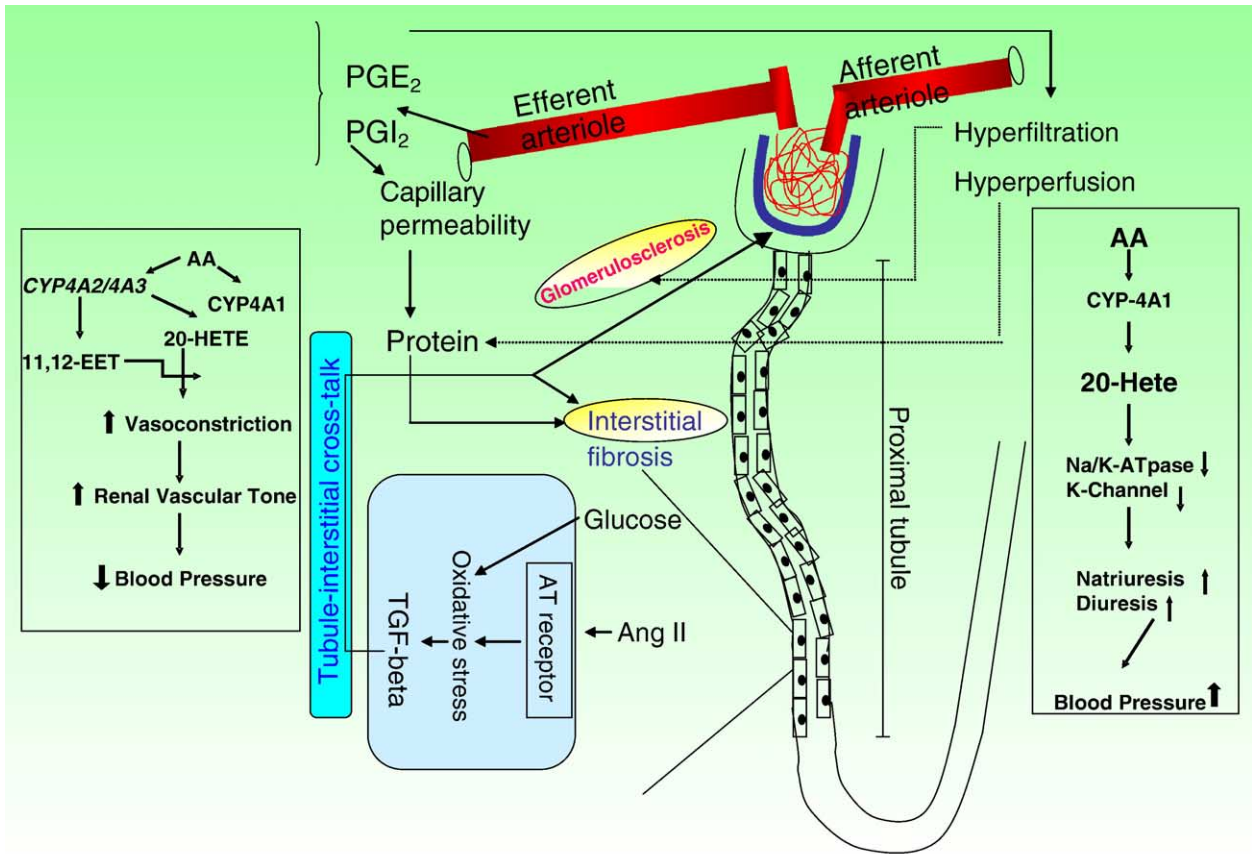


Fig. 7. Schematic presentation of HO-1 interaction to CYP450 and COX metabolites.

that the antihypertensive effect of HO enhancement may be due, in part, to blunting the vasoconstrictor action of 20-HETE formed via CYP [157,170,173].

More recently, HO inhibitors have been shown to decrease renal blood flow acutely, implying that the renal HO system supports the renal circulation via formation of CO [140,179,180]. HO-1/HO-2 and CYP450 are expressed in the renal medulla [173,181,182], in the arterial and preglomerular arteries [157], and in ECs [96,155,183,184]. Further studies have been performed in which the human HO-1 gene was delivered to SHR using a concentrated infectious viral particle [185]. Treated rats demonstrated a functional expression of the human HO-1 gene, which was associated with a significant decrease in BP in young SHR. These rats also showed a significant reduction in urinary 20-HETE, which is believed to be partially responsible for this decrease in BP [185].

### HO-arachidonic acid metabolism: Role of cyclooxygenases

COX-1 and COX-2, heme-dependent enzymes, are responsible for the conversion of AA to PGH<sub>2</sub>, which is further metabolized by thromboxane (TX) synthase, PGE synthase, and prostacyclin synthase to TXA<sub>2</sub>, PGE<sub>2</sub>, and

PGI<sub>2</sub>, respectively, and to other prostaglandins (PGF<sub>2α</sub> and PGD<sub>2</sub>) by other proteins (Fig. 8). The two COX isoforms are expressed in the rat kidney. COX-1 is constitutively expressed and has been localized to arteries and arterioles,

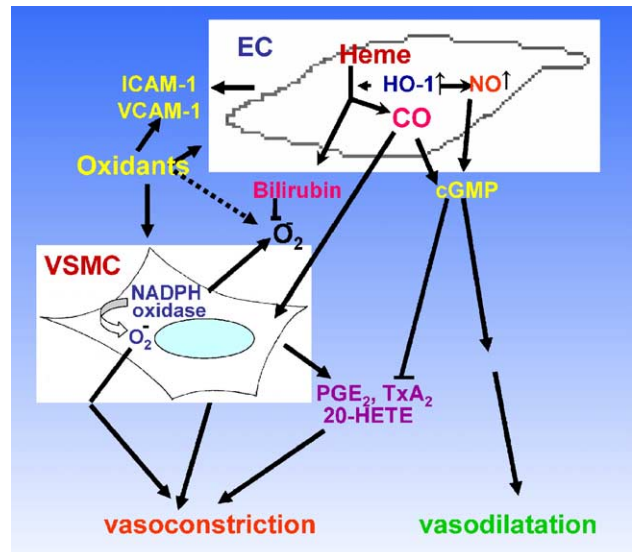


Fig. 8. Schematic presentation of interaction of HO/CO in endothelial cells and vascular smooth muscle cells.

glomeruli, and collecting ducts [186]. Although COX-2 is considered to be the inducible form, it is constitutively expressed in the TALH and in the region of the macula densa, primarily the cortical structure [187,188], where it is involved in the stimulation of renin release from the juxtaglomerular cells [189]. Studies have demonstrated that, in cells overexpressing HO-1, COX isoform expression and activity are impaired, and induction of HO-1 by SnCl<sub>2</sub> significantly reduced the expression of COX-2 in the rat kidney. Other studies have shown that eicosanoid-dependent mechanisms participate in the regulation of ion and water movement in the mammalian nephron [165]. COX-derived eicosanoids may be responsible for either prohypertensive or antihypertensive mechanisms. PGI<sub>2</sub> is a potent vasodilator and early studies have documented that endogenous PGE<sub>2</sub> production increases in response to volume depletion to help maintain GFR by dilating the afferent arteriole [190].

In the kidney, COX-2 expression is restricted to macula densa-containing segments, the cortical thick ascending limb (cTAL) [187], and, at significantly lower levels, the inner medullary collecting duct (IMCD). The level of COX-2 in these segments is regulated by dietary salt intake. In rats on a low-salt diet, cortical COX-2 increases while it decreases in rats on a high-salt diet, suggesting a role in the regulation of glomerular circulation and renin release. On the other hand, medullary COX-2 expression increased in rats on a high-salt diet and decreased in rats on a low-salt diet, suggesting a role in the regulation of salt excretion [187]. Traynor et al. [189] have recently demonstrated that COX-2 inhibition abolished the increase in renin release stimulated by low luminal NaCl concentration, supporting the role of macula densa COX-2 in the regulation of renin release. Other convincing evidence was provided by studying COX-2<sup>-/-</sup> null mice; these mice have attenuated renin expression in response to either a low-salt diet [191] or ACE inhibition [192].

COX may convert the CYP-AA metabolite, 20-HETE, to a vasoconstrictor in rat aortic rings [156,167]. Sessa et al. [160] have also showed that upregulation of HO by SnCl<sub>2</sub> resulted in corresponding decreases in renal heme content, CYP450 content, and renal thromboxane A<sub>2</sub> synthase activity in young SHR. These results also demonstrated that renal HO-1 induction via SnCl<sub>2</sub> administration affected COX-2 protein expression, leading to a marked decrease in its levels in the cortex as well as in the inner and outer medulla [154]. Haider et al. have shown that upregulation of HO activity elicits a decrease in PGE<sub>2</sub> levels in rabbit coronary microvessel ECs [193]. Several reports [187,189,191] have shown that macula densa COX-2 is involved in the regulation of renin release from the juxtaglomerular apparatus (JGA) in the afferent arteriole. These support the notion that HO activity, through regulation of heme content and CO generation, may modulate COX expression and/or activity on the molecular or biochemical levels. Whether the HO system regulates renin release, directly or indirectly via COX-2, remains to be investigated.

## HO and endothelial cell-cycle progression in diabetes

An expanding body of information has shown that progression through the mammalian cell cycle is orchestrated by distinct, multiple holoenzymes composed of catalytic subunits, cyclin-dependent kinases (cdk), whose activities depend upon a regulatory protein called cyclin [194–196]. It is well established that cdk levels remain high throughout the cell cycle. Their activity is mainly regulated by cyclin binding, with the levels of cyclin fluctuating at different stages of the cell cycle. Cell-cycle transition from G<sub>1</sub> to the S phase is controlled by three types of cyclins, D, E, and A. Different cyclins are expressed at specific stages of the cell cycle and bind to a particular cdk. In addition to the cdk-specific positive regulation of the cell cycle, cdk activities are also controlled by a new class of small proteins, the so-called cyclin kinase inhibitors (CKI); the latter bind to cyclin-cdk complexes and inhibit their kinase activity. CKI inhibitor proteins are specific for cdk4 and cdk6 [197,198]. Overexpression of these inhibitory molecules leads to cell-cycle arrest in G<sub>1</sub>.

High glucose conditions facilitate the oxidative-mediated inhibition of EC proliferation [199]. Overexpression of human HO-1 in ECs may have the potential to provide protection against a variety of agents that cause oxidative stress. The role of endogenously produced CO in the prevention of EC apoptosis has been addressed by examining the effect of conditions that affect HO activity and expression [200]. The antiapoptotic effect of HO-1-derived CO release has been shown to be mediated via the activation of p38 of mitogen-activated PKC [61,201]. Increased HO-derived CO has been shown to significantly shift, to the left, the concentration–cell death curve. This effect, however, can be reversed by the inhibition of HO [73]. These observations support the idea that the vascular HO system involves mechanisms that decrease apoptosis mediated by oxidative stress-inducing factors, including hypoxia or inflammatory molecules, such as TNF [50,51,202].

Using retrovirus-mediated human HO-1 gene transfer, it was demonstrated that HO-1 attenuates glucose-mediated cell growth arrest and apoptosis in human microvessel ECs [25]. In this study, incubation of ECs in a high-glucose medium resulted in a decrease of HO activity and a decrease in HO-1 and HO-2 proteins compared with cells exposed to low glucose or cells exposed to mannitol. As shown in Fig. 9A, apoptosis was induced in both control ECs and ECs transduced with HO-1 antisense. In contrast, cells transduced with HO-1 in the sense orientation were not affected by glucose. However, the addition of tin mesoporphyrin (SnMP), an inhibitor of HO activity, reversed the cytoprotective effect of HO-1 overexpression against the glucose (Fig. 9B).

Overexpression of HO-1 was coupled with an increase in HO activity and CO synthesis, decreased cellular heme, and acceleration in all phases of the cell cycle. The rate of the

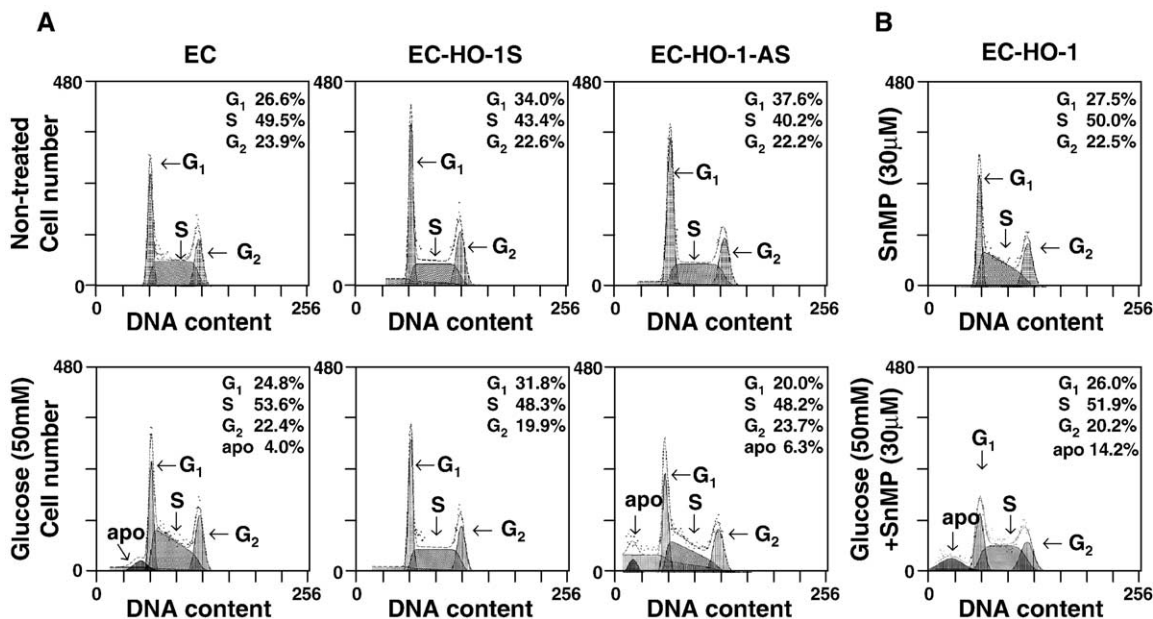


Fig. 9. The effect of high glucose on DNA distribution: (A) Control cells, nonexposed cells, and glucose-exposed control cells and cells transduced with the HO-1 sense or HO-1 antisense were stained with DAPI and analyzed by flow cytometry. Representative DNA distributions are shown. (B) Cells transduced with HO-1 and treated with SnMP while exposed to high glucose were stained with DAPI and analyzed by flow cytometry. Representative DNA distributions are shown. EC, control endothelial cells; EC-HO-1S (sense), endothelial cells transduced with the HO-1 in the sense orientation; EC-HO-1AS (antisense), endothelial cells transduced with the HO-1 in the antisense orientation [25].

cell cycle or cell birth rate was increased in cells overexpressing HO-1, but was decreased in cells underexpressing HO-1 compared with control cells. Exposure to high glucose significantly decreased cell-cycle progression in control cells and in cells underexpressing HO-1, but did not decrease cell-cycle progression in cells overexpressing HO-1. The protein levels of the cdk inhibitors, p21 and p27, were greatly affected by the level of expression of HO-1. High glucose induces p21 and p27 in control cells, but not in cells overexpressing HO-1. The addition of SnMP reversed the HO-1-mediated decrease of p21 and p27 in cells overexpressing HO-1. As seen in Fig. 10, the basal and glucose-

induced p21 levels were decreased in cells overexpressing HO-1 compared to control cells. In contrast, in HO-1-deficient cells, basal and glucose-induced p21 levels were increased. Likewise, basal and glucose-induced p27 levels were significantly reduced in cells overexpressing HO-1 and increased in HO-1-deficient cells compared to control cells. These findings identify a novel effect of HO-1 on EC growth and indicate that heme metabolism and HO-1 expression regulate signaling systems in cells exposed to high glucose, which control cell-cycle progression.

The view that HO-1 and HO-2 contribute to antiapoptotic mechanism(s) has also received support from studies on the effect of HO inducers on protection from oxidative stress-mediated cell death [95]. Another study was performed in which a retroviral vector was used to investigate the physiological significance of human HO-1 overexpression on cell-cycle progression in the presence and absence of oxidants, such as pyrrolidine dithiocarbamate [97]. This was suggested to be a potential therapeutic means for attenuating the effects of oxidative stress-causing agents [97]. The role of human HO-1 in cell-cycle progression following exposure to heme or human HO-1 gene transfer, for the identification of target genes associated with human HO-1-mediated increases in cell-cycle progression, was examined using cDNA microarray technology [102]. The results showed the upregulation of several genes associated with cell-cycle progression, including cyclin E and D; downregulation of the cdk inhibitors p21 and p27, the cdk 2, 5, and 6, and monocyte chemoattractant protein-1; and the upregulation of growth factors, including vascular endothelial growth factor (VEGF), vascular endothelial growth

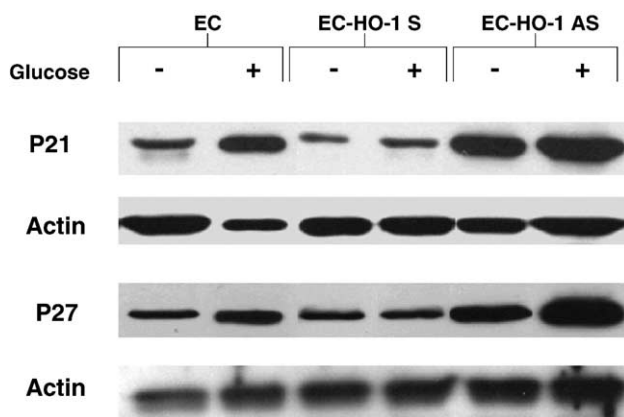


Fig. 10. Effect of high glucose on p21 and p27 protein levels in cells overexpressing (HO-1 sense) and underexpressing HO-1 (HO-1 antisense). Cells were exposed to glucose. The levels of p21 and p27 were visualized by immunoblotting with antibodies against p21 and p27. Representative blots are shown [25].

factor receptor I (VEGFRI), endothelial growth factor (EGF) and hepatic-derived growth factor (HDGF). These findings identify an array of gene responses to over-expression of human HO-1 and elucidate new aspects of human HO-1 signaling involved in cell growth.

More recently, Colombrita et al. [203] reported that expression of HO-1 in human ECs in the  $G_0/G_1$  phase, in the presence of Ang II, might be a key player in attenuating DNA damage during cell-cycle progression. Exposure of ECs to Ang II causes a complex response involving the generation of  $O_2^-$ , which may be involved in DNA damage. Thus, upregulation of HO-1 ensures the generation of bilirubin and CO in the  $G_0/G_1$  phase to counteract the Ang II-mediated oxidative DNA damage. Inducibility of HO-1 in the  $G_0/G_1$  phase is essential for protecting cells from DNA damage and ensuring cell-cycle progression.

### HO-1 and endothelial cell dysfunction in diabetes

In type 1 diabetes mellitus, insulin deficiency provokes high blood glucose levels and lipid metabolism alterations. Evolution of the disease may be associated with the development of premature micro- and macrovascular complications, the pathogenesis of which may be linked in part to oxidative stress.  $O_2^-$  is vasoconstrictive through the removal of vasodilators and the stimulation of vasoconstrictors; e.g.,  $O_2^-$  can convert NO to peroxynitrite, thereby consuming the endogenous vasodilator in the vasculature [204,205]. Additionally,  $O_2^-$  can induce assorted vasoconstrictors, such as endothelin, PDGF,  $TxA_2$ , and isoprostanes [204,205]. Reports have indicated that oxidative stress, as a result of hyperglycemia, plays an important role in the development and progression of diabetic vascular complications, such as nephropathy [206]. Hyperglycemic rats have also shown an increase in urinary 8-epi-isoprostane  $PGF_2$ ,  $O_2^-$  formation, and the number of circulating endothelial cells and fragments [26].

As seen in Fig. 11, studies from these laboratories [25,26] imply that it is the products of HO-1-mediated heme degradation (i.e., CO and bilirubin) that produce the antiapoptotic effect of HO-1. In collating the different effects of bilirubin and CO, we have hypothesized that the products of HO metabolism of heme serve as a counterbalancing influence on Ang II-mediated damage in ECs and in the vasculature by actions that include decreasing EC sloughing, inhibition of  $O_2^-$ , and the expression of inflammatory molecules (i.e., TNF, Ang II, ICAM), and a reduction in constrictor mechanisms.

In a study conducted by Koya et al. [206], diabetic glomeruli showed a 16-fold increase in the expression of HO-1 mRNA and protein. Further, they showed that through the use of vitamin E and other antioxidants, HO-1 levels are eventually normalized, indicating that antioxidants could be a potential therapeutic treatment for diabetes-related nephropathy. In addition, animals preconditioned with HO-1

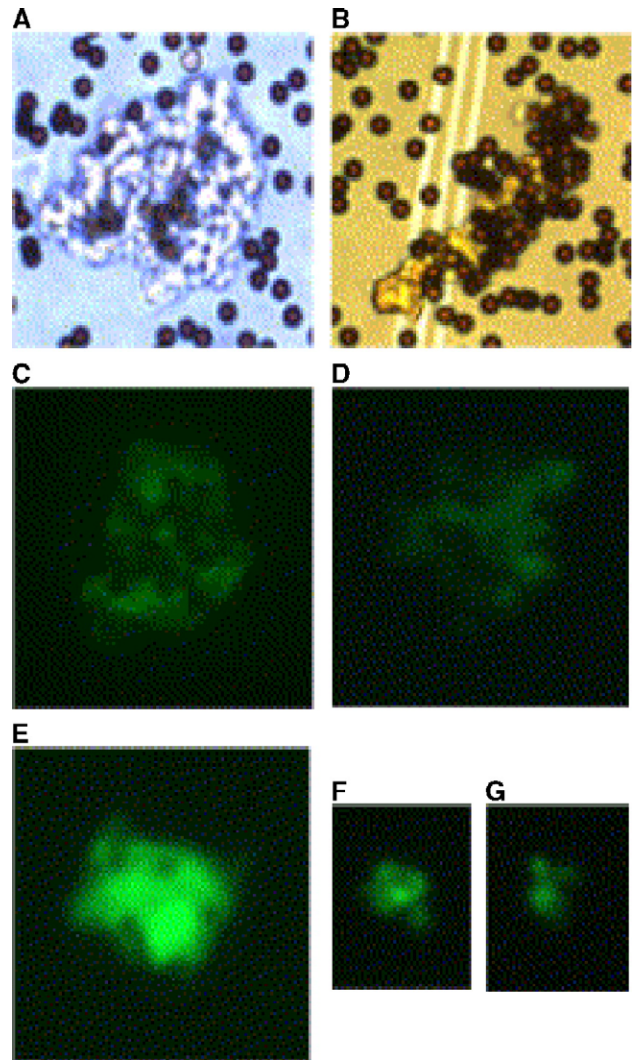


Fig. 11. Morphology of circulating endothelial cells with Dynabeads attached in STZ-induced diabetic rats. Typical circulating endothelial cells and cell debris under light microscope (A, B) or fluorescent microscope (C–G) [207].

show lower levels of  $O_2^-$  formation [25,207], vasoconstriction, and circulating ECs and fragments [70]. McClung et al. [70] concluded that HO-1 upregulation decreases neointimal vessel size associated with a reduction in  $O_2^-$  production and a decrease in CECs in balloon-injured diabetic rats, suggesting that it may have a role in the treatment of vascular restenosis.

### HO and pharmacologic/genetic interventions in cardiovascular disease

#### A. Cardiovascular drugs and HO-1 induction

A variety of drugs have been developed to reduce the complications of cardiovascular disease. The most common of these is aspirin, which has been reported to decrease platelet COX-2 levels and thus reduce cardiovascular risk.

Grosser et al. [208] have reported that aspirin increased HO-1 protein levels and activity in a dose-dependent manner in cultured ECs derived from human umbilical vein. Pretreatment of cells with aspirin or bilirubin protected ECs from hydrogen peroxide-mediated toxicity. The authors concluded that aspirin targets HO-1, via NO-dependent pathways similar to NOS blockers, such as L-NAME, preventing aspirin-dependent HO-1 induction. Oberle et al. [209] expanded these studies using the NO donor, pentaerythritol tetranitrate (PETN). They showed that the active PETN metabolite, pentaerythritol trinitrate (PETriN), increased HO-1 mRNA and protein levels. This was accompanied by an increase in enzyme activity as measured by increased CO and bilirubin production. Polte et al. [210] demonstrated that HO-1 is a cGMP-sensitive endothelial gene and established a causal relationship between HO-1 induction and endothelial protection by the cGMP/NO system. Aspirin has also been shown to increase ferritin synthesis in ECs, presumably as a result of HO-1 induction and iron release, suggesting a role in the prevention of endothelial injury during atherogenesis. Grosser and co-workers [209,211,212] have examined the mechanism of statins with respect to their antioxidation action. In cultured ECs derived from human umbilical vein, simvastatin and lovastatin increased HO-1 mRNA levels [211,213]. Increased transcriptional expression in statin-mediated HO-1 was associated with elevated HO-1 protein levels and a reduction in free radical formation. Lee et al. also reported that simvastatin activates HO-1 in VSMCs in vitro and in vivo and suggested the involvement of p38 and the p13K-Akt pathway in HO-1 induction. The authors concluded that the anti-inflammatory and antiproliferative effects of simvastatin are largely mediated through the induction of HO-1 [212].

### B. Rationale for gene transfer

Pharmacological agents such as SnMP and cobalt protoporphyrin (CoPP) can inhibit or induce HO activity significantly and effectively [13–15,127]. However, such agents can exert only transient control of HO activity. Current pharmacological agents effectively treat cardiovascular diseases such as essential vascular hypertension, but may not prevent end organ damage or influence the propensity for other diseases. Changes in CO and bilirubin formation and heme content as a result of HO-1 protein expression with genetic interventions are modest and less abrupt or volatile than those obtained after bolus administration of chemical inducers. Genetic interventions result in a steady change in HO activity and heme content, which is regulated by an increase in the rate of heme synthesis [11]. Human gene therapy for cardiovascular disease has the potential to provide important advances in therapeutic angiogenesis, myocardial protection, myocardial regeneration and repair, restenosis, prevention of bypass graft failure, and risk-factor management.

Gene transfer is a powerful tool that can be used to insert specific genes into cells otherwise deficient or that under-express the gene. Overexpression of the HO gene by targeted gene transfer has become a powerful tool for studying the role of the human HO-1 enzyme. The delivery of the human HO-1 gene to rats has been a successful approach for inducing the long-term overexpression of human HO-1. Delivery of human HO-1 into SHR has been shown not only to attenuate hypertension but also to enhance somatic body growth and cell proliferation [176]. Successful and functional HO gene transfer requires two essential elements [11]. First, the HO gene must be delivered by a safe vector, and second with the exception of HO gene delivery to ocular or cardiovascular tissue via catheter-based interventions, HO gene delivery must be site and organ specific.

Abraham et al. [6,214] described the first adenoviral construct used to deliver the human HO-1 gene. It was found that the human HO-1 gene could be introduced into rabbit ocular tissues by microinjection of a recombinant replication-deficient adenovirus human HO-1 cDNA [6,214]. Microinjection of the Adv-HHO construct into the vitreous resulted in HO-1 mRNA expression in the corneal endothelium, iris, lens, and retina. After intracameral injection, human HO-1 mRNA was detected in the corneal epithelium and endothelium, ciliary body, lens, and iris, and acted to decrease oxidant-mediated injury [6,214]. Regardless of the injection site, transfected human HO-1 mRNA was site-specific and undetectable in tissues outside the eye. The ability to transfect the human HO gene and to demonstrate its expression may offer a new therapeutic strategy for treating pathological disorders in humans.

### C. Gene transfer to correct for cardiovascular disorders

The ultimate goal in the treatment of cardiovascular disease is the timely delivery of the best therapeutic agents, which would be able to protect the heart from the deleterious effects of prolonged ischemia or the effects of repeated bouts of ischemia [215]. Ischemia and reperfusion represent major mechanisms of tissue injury and organ failure. In addition, myocardial ischemia may be asymptomatic and repeated. As a result, patients may not receive timely treatments [215]. Pachori et al. [216] have developed a preemptive strategy for tissue protection using an adeno-associated vector system, containing erythropoietin hypoxia response elements, for ischemia-regulated expression of the therapeutic human HO-1 gene. They demonstrated that a single administration of this vector several weeks in advance of ischemia/reperfusion injury to the heart produced a rapid and timely induction of human HO-1 during ischemia, which resulted in a dramatic reduction in tissue damage. In addition, they also showed that overexpression of the therapeutic transgene prevented long-term pathological tissue remodeling and normalized tissue function. A similar study done by Coito et al. [217], with regards to ischemia-

induced myocardial injury, demonstrated that rats having undergone successful HO-1 gene administration had a dramatic reduction in left ventricular myocardial infarction after coronary artery ligation and release. In addition to the reduction in infarct size, there was a decrease in myocardial lipid peroxidation, proapoptotic Bax, and proinflammatory interleukin-1 beta (IL-1 $\beta$ ) protein abundance, as well as an increase in antiapoptotic Bcl-2 protein levels. These data demonstrate that HO-1 gene transfer produces therapeutic, cardioprotective benefits by reducing oxidative stress and associated inflammation and cell death.

According to Vassalli et al. [218], current methods of heart transplantation are limited by incomplete effectiveness, significant toxicity, and failure to prevent chronic rejection. Genetic manipulation of the donor heart at the time of removal offers a unique opportunity to introduce a therapeutic molecule within the graft itself while minimizing systemic effects. Cytoprotective approaches, including HO-1 gene transfer, have reduced ischemia/reperfusion injury and delayed cardiac allograft rejection in small animals. However, despite major experimental advances, gene therapies for heart transplantation have not yet entered the clinical arena due to unanswered questions regarding the most suitable vector, the best gene, and safety issues. Gene therapy is being further considered for allograft repair. Braudeau et al. [219] analyzed the effects of specific overexpression of HO-1 following adenovirus-mediated HO-1 gene transfer in an acute cardiac allograft rejection model. These results indicate that HO-1 overexpression prolongs the survival of vascularized allografts by promoting tolerogenic mechanisms acting on allogeneic cellular immune responses.

Others such as Tang et al. [220] believe, however, that the constitutive overexpression of human HO-1 may lead to unwanted side effects. As a result, they designed a hypoxia-regulated human HO-1 gene therapy system that can be switched on and off. Their vigilant plasmid system is composed of a myosin light-chain-2v promoter and a gene switch that is based on an oxygen-dependent degradation domain from the hypoxia-inducible factor-1-alpha (HIF-1 $\alpha$ ). This vector can sense ischemia and switch on the human HO-1 gene system, specifically in the heart. Subsequently, its expression can then turn off, after sensing that the tissue is adequately oxygenated, once proper blood flow is restored to the heart. Hearts from HO-1 gene therapy-treated mice showed improved recovery of contractile and diastolic performance after myocardial infarction compared with saline controls. This study documents the therapeutic potential of vigilant plasmid-mediated human HO-1 gene transfer, which may provide cardiac-specific protection from repeated bouts of ischemic injury.

#### D. Gene transfer and hypertension

Gene transfer for the management of hypertension has considerable clinical potential, especially if the gene is

delivered to the vascular system before hypertension becomes sustained. Goodman et al. [185] and others [10] performed a study using a retroviral vector in which they examined the effects of human HO-1 gene delivery on cellular heme in renal tissues. Analysis indicated an increase in human HO-1 distribution and activity throughout the renal tissue, which was accompanied by a significant reduction in blood pressure. Sabaawy et al. [176] performed a similar experiment in which they gave a single intracardiac injection of concentrated infectious viral particles expressing human HO-1 to SHR. These results, which were similar to those of Goodman et al., demonstrated functional expression of human HO-1 and attenuation of development of hypertension.

It was recently shown that delivery of the HO-1 gene, in sense or antisense orientation, alters HO-1 expression and subsequently regulates BP [10]. A retrovirus carrying the human HO-1 gene in its sense orientation (LSN-HHO-1) or the rat HO-1 in its antisense orientation (LSN-RHO-AS) was administered to newborn Sprague-Dawley (SD) rats. Two months later the rats were exposed to Ang II. Rats exposed to the human HO-1 gene in its antisense orientation demonstrated decreased HO-1 expression and showed an increase in BP after exposure to Ang II. In contrast, rats given the human HO-1 gene in its sense orientation demonstrated human HO-1 expression and a decrease in BP after being exposed to Ang II. These data demonstrate that overexpression of HO-1 brings about a reduction in pressor responsiveness to Ang II, which is most likely due to the increased generation of an HO-1 product, presumably CO, with the ability to inhibit vascular reactivity to constrictor stimuli.

#### E. Gene transfer and diabetes

Diabetes is another example of a metabolic disturbance in which HO-1 gene transfer in the experimental animal has been shown to be beneficial. Although oxidative stress is induced under diabetic conditions and causes various forms of tissue damage, acute diabetes does not result in an increase HO-1 protein [25,26]. Abraham et al. [26] compared the basal levels of HO-1 and HO-2 proteins, as well as the effect of heme on these levels, in aorta from control and hyperglycemic rats using immunohistochemistry. In hyperglycemic rats, HO-1 immunoreactivity was reduced in the intima, but not in the media or adventitia. In contrast to control rats, HO-1 immunoreactivity in the intima and media was less intense in hyperglycemic rats treated with heme (Figs. 12C and D; left panel). HO-2 immunoreactivity was evident in the intima, media, and adventitia, and was not affected by heme or hyperglycemia. Moderate HO-1 immunoreactivity was observed in the intima, media, and adventitia of the aorta from control rats. Heme administration resulted in strong immunoreactivity in the intima (thin arrow) and in the media (thick arrow) (Fig. 12B, right upper panel). This study and other suggested that

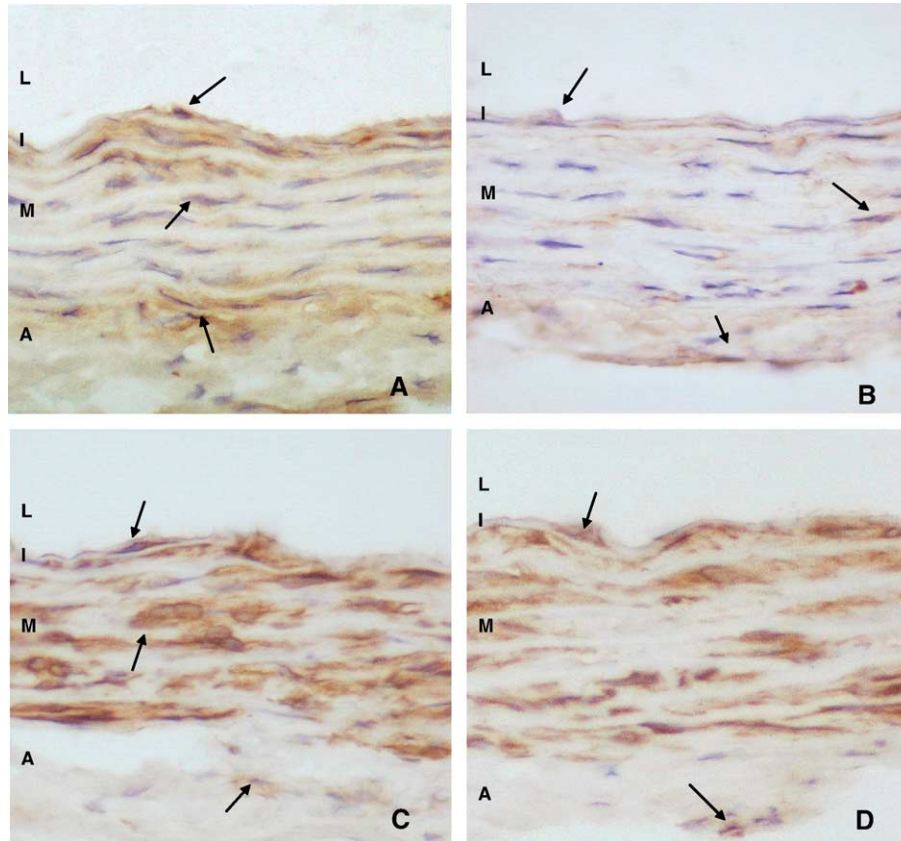


Fig. 12. HO-1 immunohistochemistry in aorta from control (A), heme-treated (B), STZ-treated (C), and STZ plus heme-treated rats (D). Heme was given prior to sacrifice. In STZ animals, HO-1 staining of intima was decreased with respect to controls whereas in the media, as in controls, some focal areas of positivity are present. Administration of heme restored HO-1 staining. Immunopositivity was a dark brown stain in the cytoplasm of cells. Nuclei are counterstained with hematoxylin [26].

delivery of the human HO-1 gene, or HO-1 preconditioning, may provide cytoprotection to the endothelium [26].

Abraham et al. [26] also assessed the functional expression of human HO-1 gene transfer as determined by the levels of human HO-1 protein in the aorta of rats receiving LSN-HHO-1 compared to controls receiving empty retrovirus vector (LSXN) [26]. Western blot analysis of aorta obtained from LSN-HHO-1 rats demonstrated human HO-1 protein expression [26]. In contrast, no human HO-1 protein was expressed in aorta from rats injected with the empty retrovirus vector. These data confirm the functional expression of the human HO-1 gene on HO-1 protein levels after intracardiac delivery of LSN-HHO-1. More importantly, neither rat HO-2 nor actin were affected by expression of the human HO-1 gene as indicated by comparable expression of HO-2 and actin in rats receiving human HO-1 gene transfer or in LSXN-treated rats. The effect of hyperglycemia on HO activity in rats overexpressing (LSN-HHO-1) and underexpressing HO-1 (LSN-rHO-1 AS) was assessed [26]. Streptozotocin (STZ)-induced hyperglycemia resulted in significantly reduced HO activity in both control rats and rats transduced with LSN-rHO-1-AS (Fig. 13).

The effect of HO overexpression and underexpression on the generation of 8-epi-isoprostane  $\text{PGF}_{2\alpha}$ , which is

considered as a reliable index of oxidative stress status in vivo in normal and hyperglycemic rats, was also studied [26]. Urinary excretion of 8-epi-isoprostane  $\text{PGF}_{2\alpha}$  was increased in hyperglycemic rats compared to control rats, but not in rats overexpressing human HO-1 when compared to controls. Rats overexpressing HO-1 (LSN-HHO-1) showed a significant decrease in urinary excretion of 8-epi-isoprostane  $\text{PGF}_{2\alpha}$ . In contrast, rats underexpressing HO-1 excreted higher levels of 8-epi-isoprostane  $\text{PGF}_{2\alpha}$ , which were further increased as a result of hyperglycemia. Underexpression of HO-1 exacerbated excretion of 8-epi-isoprostane  $\text{PGF}_{2\alpha}$ , increasing its output significantly in LSN-rHO-1 AS rats. The mechanism by which human HO-1 gene transfer into rats exerts antioxidant effects was further studied by measuring  $\text{O}_2^-$  in control and diabetic rat vessels overexpressing and underexpressing HO activity (Fig. 13). Production of  $\text{O}_2^-$  was increased in diabetic rats compared to controls. The report also examined whether the preventive vascular protection induced by human HO-1 gene transfer would decrease vascular  $\text{O}_2^-$ . HO-1 gene transfer attenuated the hyperglycemia-mediated increase in  $\text{O}_2^-$ . In contrast, diminished HO-1 expression potentiated the hyperglycemia-mediated increase in  $\text{O}_2^-$  seen in transgenic HO-1 AS rats. These results provide additional



evidence that pharmacological induction of HO-1 or delivery of the HO-1 gene to the vascular system can provide vascular protection and subserves an antioxidant defense mechanism against pathological conditions, in which decreasing  $O_2^-$  formation is an important goal.

The effect of hyperglycemia on EC damage and sloughing into the circulation in control and hyperglycemic rats overexpressing or underexpressing HO-1 was also examined [26]. The basal levels of circulating EC in HO-1 transgenic and control rats before the onset of diabetes were not substantially different, but were significantly elevated in transgenic HO-1-AS rats as a result of diminished HO activity. As seen in Fig. 14, the number of circulating EC in blood obtained from control rats was low.

However, a large increase in the number of circulating EC was observed in blood obtained from hyperglycemic rats, similar to that seen in other pathologic circumstances. The number of circulating EC in hyperglycemic rats overexpressing HO-1 was decreased. These results demonstrate that the vascular system may develop an adaptive response to hyperglycemia-mediated stress as a result of transduction of the HO-1 gene and the decreased detachment of EC. In contrast, the number of circulating endothelial cells in hyperglycemic rats underexpressing HO-1 was significantly higher than that in control hyperglycemic rats, suggesting that the extent and severity of the increase in circulating endothelial cells were not predetermined at the onset of diabetes, but could be modified by the level of HO-1 gene

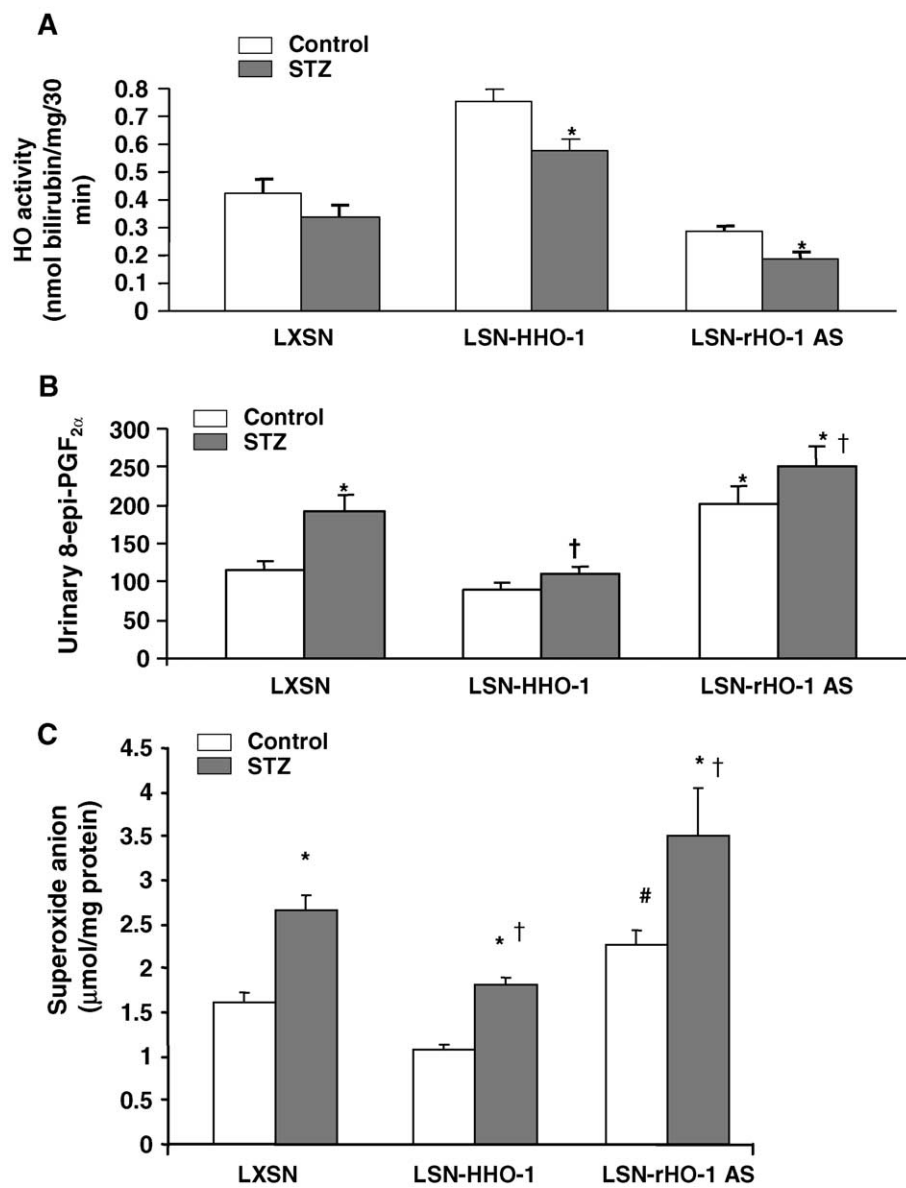


Fig. 13. Effect of hyperglycemia on (A) HO activity; (B) rats transduced with in aorta from rats transduced with LXSN, LSN-HHO-1, and LSN-rHO-1 AS. HO activity and urinary 8-epi-isoprostane  $PGF_{2\alpha}$  were measured as described in the text. \* $p < 0.05$  vs. the corresponding control (not treated with STZ); † $p < 0.05$  vs. STZ-treated LXSN transduced rats. (C) Aortic  $O_2^-$  production in STZ rats transduced with LXSN, LSN-HHO-1, and LSN-rHO-1 AS.  $O_2^-$  production was measured as described in the text. \* $p < 0.05$  vs. the corresponding control (not treated with STZ); † $p < 0.05$  vs. STZ-treated LXSN transduced rats.

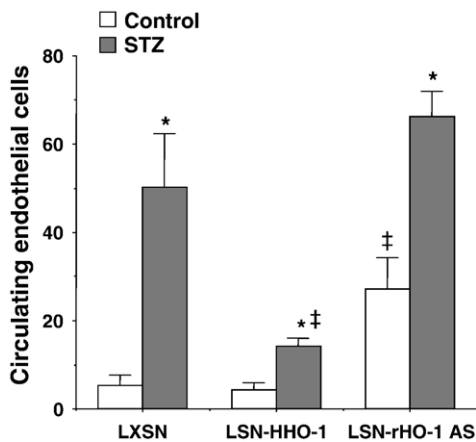


Fig. 14. Effect of hyperglycemia on the number of circulating endothelial cells. Diabetic rats demonstrated a significant increase in the number of circulating endothelial cells when compared to nondiabetic control rats. Furthermore, diabetic rats preconditioned with the HO-1 gene showed a significant decrease in the number of circulating endothelial cells when compared to untreated diabetic rats. The upregulation of HO-1 has a cytoprotective effect, which prevents endothelial cell sloughing [26].

expression. These data demonstrate that HO-1 gene transfer into normal rats at an early stage during the development of the vascular system may attenuate cardiovascular complications during the development of diabetes [26,207].

#### F. Gene targeting

Site- and tissue-specific transcriptional regulating elements represent an alternative strategy for restricting adenoviral transgene expression to specific cell lineages or tissues *in vivo* [221,222]. This strategy has been recently applied using a SMC-specific SM22 $\alpha$  promoter to direct expression of recombinant gene products to vascular or visceral SMCs *in vivo* [222]. A few examples of cell lineage-specific promoters are listed in Table 1.

The mTALH is situated in a site of markedly diminished oxygen tension and is highly vulnerable to ischemic insult involving Ang II-mediated increases in COX-2 activity. Quan et al. [27] demonstrated that the site-specific delivery of HO-1 to renal structures, specifically the mTALH, in SHR, using the Na<sup>+</sup>-K<sup>+</sup>-Cl<sup>-</sup> cotransporter (NKCC2 promoter), normalized BP and provided protection to the mTALH against oxidative stress caused by Ang II. Western blot and RT-PCR revealed that human HO-1 was selectively expressed in primary cultured TALH cells following infection with Ad-NKCC2-HO-1. In TALH cells infected with Ad-NKCC2-HO-1, Ang II-stimulated PGE<sub>2</sub> levels were significantly reduced. Ang II caused a marked decrease in reduced glutathione (GSH) levels and this decrease was greatly attenuated in TALH cells upregulated with HO-1. This is the first demonstration that HO-1 overexpression increases reduced GSH [27]. Others have shown that upregulation of HO-1 may decrease cellular heme availability for the synthesis gp91 (phox) and related NOX oxidase and the generation of O<sub>2</sub><sup>-</sup> [227].

#### Concluding remarks

The heme oxygenase (HO) system plays an important role in many aspects of human physiology, as well as in pathological circumstances associated with myocardial ischemia/reperfusion, hypertension, cardiomyopathy, organ transplantation, endotoxemia, and pulmonary disorders among others. Active agents in these situations include not only heme itself, but its metabolic products, CO, and bile pigments as well as those agents derived from the wide array of genetic and metabolic processes which respond when heme metabolism is perturbed [102]. The use of pharmacological and genetic interventions for regulating heme oxygenase has already provided important new insights into the relation of the heme–HO system to biological and pathological events and offers the potential for development of new therapeutic strategies directed against recalcitrant disease processes. It is, for example, possible to envisage the use of a single drug or gene intervention using site-specific expression to induce long-term prophylaxis against certain pathological cardiovascular events or to promote and enhance repair processes in individuals who have experienced, or are at high risk for, cardiac injury. The use of stable gene integration may also lead to the development of more effective, perhaps long-lasting, therapies which can moderate other chronic diseases.

The HO system is on the one hand a seemingly simple biochemical axis and on the other, the hub of a complex of coupled processes whose actions and products exert a wide array of diverse and potent metabolic effects. Down regulating the HO system by pharmacological or genetic means has already led to new approaches for managing old clinical problems [15] and up-regulating the system offers even more promise for new approaches to experimental work in cell biology and new methods for moderating some of the consequences of clinical disorders. The use of pharmacological and genetic interventions for upregulation of HO-1 in the managements of cardiovascular diseases appears to be especially promising. In the future, it may be possible to utilize a single administration of a drug or gene therapy using site-specific expression of HO-1/HO-2 to promote long-term prophylaxis against secondary coronary

Table 1

| Promoter  | Specific cell lineage          | Transgene    |
|---|--------------------------------|--------------|
| SM22 $\alpha$ early marker gene for SMCs              | SMCs [222]                     | HO/CYP/COX-2 |
| GGT ( $\alpha$ -glutamyl transpeptidase)              | Proximal tubules [223]         | HO/CYP/COX-2 |
| KAP-HAGT (kidney androgen-regulated protein promoter) | Proximal tubules [224]         | HO/CYP4AS    |
| <i>Tie</i> (tyrosine kinase receptor)                 | Endothelium/bloodvessels [225] | HO/CYP4AS    |
| <i>NKcc2/Slc12 a1</i> (Na-K-Cl cotransporter)         | TALH [226]                     | COX-2/HO     |

events and to promote myocardial repair in patients who have experienced an infarct as well as in those at high risk of myocardial injury. In addition, using stable gene integration, it may be possible to develop more effective and long-sustained therapies for chronic disorders such as diabetes and other metabolic diseases. These pharmacologic or genetic strategies to regulate the heme–HO system could, as we have earlier suggested [13], open up new therapeutic approaches for the effective management of a number of clinical disorders.

## Acknowledgments

This work was supported by NIH Grants HL55601 and HL34300 and Philip Morris Managements Group Inc. (N.G.A.) and gifts from the Ablon, Lang, and Renfield Foundations to The Rockefeller University (A.K.). We thank Dr. Saadet Türkseven for her outstanding computer skills and Dr. George Drummond for his helpful review of the manuscript.

## References

- [1] Abraham, N. G.; Drummond, G. S.; Lutton, J. D.; Kappas, A. The biological significance and physiological role of heme oxygenase. *Cell. Physiol. Biochem.* **6**:129–168; 1996.
- [2] Cruse, I.; Maines, M. D. Evidence suggesting that the two forms of heme oxygenase are products of different genes. *J. Biol. Chem.* **263**:3348–3353; 1988.
- [3] Huang, T. J.; Jr. McCoubrey, W. K.; Maines, M. D. Heme oxygenase-2 interaction with metalloporphyrins: function of heme regulatory motifs. *Antioxid. Redox. Signal.* **3**:685–696; 2001.
- [4] Scapagnini, G.; D'Agata, V.; Calabrese, V.; Pascale, A.; Colombrina, C.; Alkon, D.; Cavallaro, S. Gene expression profiles of heme oxygenase isoforms in the rat brain. *Brain Res.* **954**:51–59; 2002.
- [5] Ishikawa, K.; Takeuchi, N.; Takahashi, S.; Matera, K. M.; Sato, M.; Shibahara, S.; Rousseau, D. L.; Ikeda-Saito, M.; Yoshida, T. Heme oxygenase-2. Properties of the heme complex of the purified tryptic fragment of recombinant human heme oxygenase-2. *J. Biol. Chem.* **270**:6345–6350; 1995.
- [6] Abraham, N. G.; da-Silva, J. L.; Lavrovsky, Y.; Stoltz, R. A.; Kappas, A.; Dunn, M. W.; Schwartzman, M. L. Adenovirus-mediated heme oxygenase-1 gene transfer into rabbit ocular tissues. *Invest. Ophthalmol. Vis. Sci.* **36**:2202–2210; 1995.
- [7] Abraham, N. G.; Lavrovsky, Y.; Schwartzman, M. L.; Stoltz, R. A.; Levere, R. D.; Gerritsen, M. E.; Shibahara, S.; Kappas, A. Transfection of the human heme oxygenase gene into rabbit coronary microvessel endothelial cells: protective effect against heme and hemoglobin toxicity. *Proc. Natl. Acad. Sci. USA* **92**:6798–6802; 1995.
- [8] Abraham, N. G. Adenovirus-mediated heme oxygenase gene transfer into human hematopoietic CD34<sup>+</sup> stem cells. *Exp. Hematol.* **24** (9):1053; 1996.
- [9] Abraham, N. G.; Jiang, S.; Yang, L.; Zand, B. A.; Laniado-Schwartzman, M.; Marji, J.; Drummond, G. S.; Kappas, A. Adenoviral vector-mediated transfer of human heme oxygenase in rats decreases renal heme-dependent arachidonic acid epoxygenase activity. *J. Pharmacol. Exp. Ther.* **293**:494–500; 2000.
- [10] Yang, L.; Quan, S.; Nasjletti, A.; Laniado-Schwartzman, M.; Abraham, N. G. Heme oxygenase-1 gene expression modulates angiotensin II-induced increase in blood pressure. *Hypertension* **43**:1221–1226; 2004.
- [11] Abraham, N. G. Therapeutic applications of human heme oxygenase gene transfer and gene therapy. *Curr. Pharm. Des.* **9**: 2513–2524; 2003.
- [12] Dennerly, P. A.; Spitz, D. R.; Yang, G.; Tatarov, A.; Lee, C. S.; Shegog, M. L.; Poss, K. D. Oxygen toxicity and iron accumulation in the lungs of mice lacking heme oxygenase-2. *J. Clin. Invest.* **101**: 1001–1011; 1998.
- [13] Kappas, A.; Drummond, G. Control of heme metabolism with synthetic metalloporphyrins. *J. Clin. Invest.* **77**:335–339; 1986.
- [14] Drummond, G. S.; Kappas, A. The cytochrome P-450-depleted animal: an experimental model for in vivo studies in chemical biology. *Proc. Natl. Acad. Sci. USA* **79**:2384–2388; 1982.
- [15] Kappas, A. A method for interdicting the development of severe jaundice in newborns by inhibiting the production of bilirubin. *Pediatrics* **113**:119–123; 2004.
- [16] Vile, G. F.; Basu-Modak, S.; Wlatner, C.; Tyrrell, R. M. Heme oxygenase 1 mediates an adaptive response to oxidative stress in human skin fibroblasts. *Proc. Natl. Acad. Sci. USA* **91**:2607–2610; 1994.
- [17] Wagener, F. A.; da Silva, J. L.; Farley, T.; de Witte, T.; Kappas, A.; Abraham, N. G. Differential effects of heme oxygenase isoforms on heme mediation of endothelial intracellular adhesion molecule 1 expression. *J. Pharmacol. Exp. Ther.* **291**:416–423; 1999.
- [18] Balla, G.; Jacob, H. S.; Balla, J.; Rosenberg, M.; Nath, K.; Apple, F.; Easton, J. W.; Vercellotti, G. M. Ferritin: a cyto-protective antioxidant strategm of endothelium. *J. Biol. Chem.* **267**:18148–18153; 1992.
- [19] Hayashi, S.; Takamiya, R.; Yamaguchi, T.; Matsumoto, K.; Tojo, S. J.; Tamatani, T.; Kitajima, M.; Makino, N.; Ishimura, Y.; Suematsu, M. Induction of heme oxygenase-1 suppresses venular leukocyte adhesion elicited by oxidative stress: role of bilirubin generated by the enzyme. *Circ. Res.* **85**:663–671; 1999.
- [20] Platt, J. L.; Nath, K. A. Heme oxygenase: protective gene or Trojan horse. *Nat. Med.* **4**:1364–1365; 1998.
- [21] Nath, K. A.; Haggard, J. J.; Croatt, A. J.; Grande, J. P.; Poss, K. D.; Alam, J. The indispensability of heme oxygenase-1 in protecting against acute heme protein-induced toxicity in vivo. *Am. J. Pathol.* **156**:1527–1535; 2000.
- [22] Stocker, R.; Yamamoto, Y.; McDonagh, A. F.; Glazer, A. N.; Ames, B. N. Bilirubin is an antioxidant of possible physiological importance. *Science* **235**:1043–1046; 1987.
- [23] Stocker, R.; McDonagh, A. F.; Glazer, A. N.; Ames, B. N. Antioxidant activities of bile pigments: biliverdin and bilirubin. *Methods Enzymol.* **186**:301–309; 1990.
- [24] Wiesel, P.; Patel, A. P.; DiFonzo, N.; Marria, P. B.; Sim, C. U.; Pellacani, A.; Maemura, K.; LeBlanc, B. W.; Marino, K.; Doerschuk, C. M.; Yet, S. F.; Lee, M. E.; Perrella, M. A. Endotoxin-induced mortality is related to increased oxidative stress and end-organ dysfunction, not refractory hypotension, in heme oxygenase-1-deficient mice. *Circulation* **102**:3015–3022; 2000.
- [25] Abraham, N. G.; Kushida, T.; McClung, J.; Weiss, M.; Quan, S.; Lafaro, R.; Darzynkiewicz, Z.; Wolin, M. Heme oxygenase-1 attenuates glucose-mediated cell growth arrest and apoptosis in human microvessel endothelial cells. *Circ. Res.* **93**:507–514; 2003.
- [26] Abraham, N. G.; Rezzani, R.; Rodella, L.; Kruger, A.; Taller, D.; Li, V. G.; Goodman, A. I.; Kappas, A. Overexpression of human heme oxygenase-1 attenuates endothelial cell sloughing in experimental diabetes. *J. Am. Physiol. Heart Circ. Physiol.* **287**:H2468–H2477; 2004.
- [27] Quan, S.; Yang, L.; Shenouda, S.; Schwartzman, M. L.; Nasjletti,

- A.; Goodman, A. I.; Abraham, N. G. Expression of human heme oxygenase-1 in the thick ascending limb attenuates angiotensin II-mediated increase in oxidative injury. *Kidney Int.* **65**:1628–1639; 2004.
- [28] Otterbein, L. E.; Kolls, J. K.; Mantell, L. L.; Cook, J. L.; Alam, J.; Choi, A. M. Exogenous administration of heme oxygenase-1 by gene transfer provides protection against hyperoxia-induced lung injury. *J. Clin. Invest.* **103**:1047–1054; 1999.
- [29] Leffler, C. W.; Nasjletti, A.; Yu, C.; Johnson, R. A.; Fedinec, A. L.; Walker, N. Carbon monoxide and cerebral microvascular tone in newborn pigs. *Am. J. Physiol.* **276**:H1641–H1646; 1999.
- [30] Leffler, C. W.; Nasjletti, A.; Johnson, R. A.; Fedinec, A. L. Contributions of prostacyclin and nitric oxide to carbon monoxide-induced cerebrovascular dilation in piglets. *Am. J. Physiol. Heart Circ. Physiol.* **280**:H1490–H1495; 2001.
- [31] Zhang, F.; Kaide, J.; Wei, Y.; Jiang, H.; Yu, C.; Balazy, M.; Abraham, N. G.; Wang, W.; Nasjletti, A. Carbon monoxide produced by isolated arterioles attenuates pressure-induced vasoconstriction. *Am. J. Physiol. Heart Circ. Physiol.* **281**:H350–H358; 2001.
- [32] Kaide, J.-I.; Zhang, F.; Wei, Y.; Jiang, H.; Yu, C.; Wang, W. H.; Balazy, M.; Abraham, N. G.; Nasjletti, A. Carbon monoxide of vascular origin attenuates the sensitivity of renal arterial vessels to vasoconstrictors. *J. Clin. Invest.* **107**:1163–1171; 2001.
- [33] Kaide, J.; Zhang, F.; Wei, Y.; Wang, W.; Gopal, V. R.; Falck, J. R.; Laniado-Schwartzman, M.; Nasjletti, A. Vascular CO counterbalances the sensitizing influence of 20-HETE on agonist-induced vasoconstriction. *Hypertension* **44**:210–216; 2004.
- [34] Kaide, J.; Wang, M. H.; Wang, J. S.; Zhang, F.; Gopal, V. R.; Falck, J. R.; Nasjletti, A.; Laniado-Schwartzman, M. Transfection of CYP4A1 cDNA increases vascular reactivity in renal interlobar arteries. *Am. J. Physiol. Renal Physiol.* **284**:F51–F56; 2003.
- [35] Perrella, M. A.; Yet, S. F. Role of heme oxygenase-1 in cardiovascular function. *Curr. Pharm. Des.* **9**:2479–2487; 2003.
- [36] Ferris, C. D.; Jaffrey, S. R.; Sawa, A.; Takahashi, M.; Brady, S. D.; Barrow, R. K.; Tysoe, S. A.; Wolosker, H.; Baranano, D. E.; Dore, S.; Poss, K. D.; Snyder, S. H. Haem oxygenase-1 prevents cell death by regulating cellular iron. *Nat. Cell Biol.* **1**:152–157; 1999.
- [37] Conners, M. S.; Stoltz, R. A.; Davis, K. L.; Dunn, M. W.; Abraham, N. G.; Levere, R. D. A closed eye contact lens model of corneal inflammation. Part 2. Inhibition of cytochrome P450 arachidonic acid metabolism alleviates inflammatory sequelae. *Invest. Ophthalmol. Vis. Sci.* **36**:841–850; 1995.
- [38] Willis, D.; Moore, A. R.; Frederick, R.; Willoughby, D. A. Heme oxygenase: a novel target for the modulation of the inflammatory response. *Nat. Med.* **2**:87–90; 1996.
- [39] Laniado-Schwartzman, M.; Conners, M. S.; Dunn, M. W.; Levere, R. D.; Kappas, A.; Abraham, N. G. Heme oxygenase induction with attenuation of experimentally-induced corneal inflammation. *Biochem. Pharmacol.* **53**:1069–1075; 1997.
- [40] Juan, S. H.; Lee, T. S.; Tseng, K. W.; Liou, J. Y.; Shyue, S. K.; Wu, K. K.; Chau, L. Y. Adenovirus-mediated heme oxygenase-1 gene transfer inhibits the development of atherosclerosis in apolipoprotein E-deficient mice. *Circulation* **104**:1519–1525; 2001.
- [41] Ishikawa, K.; Navab, M.; Leitinger, N.; Fogelman, A. M.; Lusis, A. J. Induction of heme oxygenase-1 inhibits the monocyte transmigration induced by mildly oxidized LDL. *J. Clin. Invest.* **100**:1209–1216; 1997.
- [42] Wang, L. J.; Lee, T. S.; Lee, F. Y.; Pai, R. C.; Chau, L. Y. Expression of heme oxygenase-1 in atherosclerotic lesions. *Am. J. Pathol.* **152**:711–720; 1998.
- [43] Pileggi, A.; Molano, R. D.; Berney, T.; Cattani, P.; Vizzardelli, C.; Oliver, R.; Fraker, C.; Ricordi, C.; Pastori, R. L.; Bach, F. H.; Inverardi, L. Heme oxygenase-1 induction in islet cells results in protection from apoptosis and improved in vivo function after transplantation. *Diabetes* **50**:1983–1991; 2001.
- [44] Hashiba, T.; Suzuki, M.; Nagashima, Y.; Suzuki, S.; Inoue, S.; Tsuburai, T.; Matsuse, T.; Ishigatubo, Y. Adenovirus-mediated transfer of heme oxygenase-1 cDNA attenuates severe lung injury induced by the influenza virus in mice. *Gene Ther.* **8**:1499–1507; 2001.
- [45] Amersi, F.; Buelow, R.; Kato, H.; Ke, B.; Coito, A. J.; Shen, X. D.; Zhao, D.; Zaky, J.; Melinek, J.; Lassman, C. R.; Kolls, J. K.; Alam, J.; Ritter, T.; Volk, H. D.; Farmer, D. G.; Ghobrial, R. M.; Busuttill, R. W.; Kupiec-Weglinski, J. W. Upregulation of heme oxygenase-1 protects genetically fat Zucker rat livers from ischemia/reperfusion injury. *J. Clin. Invest.* **104**:1631–1639; 1999.
- [46] Guo, Y.; Stein, A. B.; Wu, W. J.; Tan, W.; Zhu, X.; Li, Q. H.; Dawn, B.; Motterlini, R.; Bolli, R. Administration of a CO-releasing molecule at the time of reperfusion reduces infarct size in vivo. *Am. J. Physiol. Heart Circ. Physiol.* **286**:H1649–H1653; 2004.
- [47] Yet, S. F.; Tian, R.; Layne, M. D.; Wang, Z. Y.; Maemura, K.; Solovyeva, M.; Ith, B.; Melo, L. G.; Zhang, L.; Ingwall, J. S.; Dzau, V. J.; Lee, M. E.; Perrella, M. A. Cardiac-specific expression of heme oxygenase-1 protects against ischemia and reperfusion injury in transgenic mice. *Circ. Res.* **89**:168–173; 2001.
- [48] Abraham, N. G.; Friedland, M. L.; Levere, R. D. Heme metabolism in hepatic and erythroid cells. In: Brown, E. (Ed.), *Progress in Hematology*. Grune and Stratton, New York, pp. 75–130; 1983.
- [49] Schacter, B. A. Heme catabolism by heme oxygenase: physiology, regulation, and mechanism of action. *Semin. Hematol.* **25**:349–369; 1988.
- [50] Kushida, T.; LiVolti, G.; Goodman, A. I.; Abraham, N. G. TNF-alpha-mediated cell death is attenuated by retrovirus delivery of human heme oxygenase-1 gene into human microvessel endothelial cells. *Transplant. Proc.* **34**:2973–2978; 2002.
- [51] Kushida, T.; Li, V. G.; Quan, S.; Goodman, A.; Abraham, N. G. Role of human heme oxygenase-1 in attenuating TNF-alpha-mediated inflammation injury in endothelial cells. *J. Cell. Biochem.* **87**:377–385; 2002.
- [52] Stocker, R.; Glazer, A. N.; Ames, B. N. Antioxidant activity of albumin-bound bilirubin. *Proc. Natl. Acad. Sci. USA* **84**:5918–5922; 1987.
- [53] Frei, B.; Stocker, R.; Ames, B. N. Antioxidant defenses and lipid peroxidation in human blood plasma. *Proc. Natl. Acad. Sci. USA* **85**:9748–9752; 1988.
- [54] Neuzil, J.; Stocker, R. Free and albumin-bound bilirubin are efficient co-antioxidants for alpha-tocopherol, inhibiting plasma and low density lipoprotein lipid peroxidation. *J. Biol. Chem.* **269**:16712–16719; 1994.
- [55] Wu, T. W.; Wu, J.; Li, R. K.; Mickle, D.; Carey, D. Albumin-bound bilirubins protect human ventricular myocytes against oxyradical damage. *Biochem. Cell Biol.* **69**:683–688; 1991.
- [56] Dore, S.; Takahashi, M.; Ferris, C. D.; Zakhary, R.; Hester, L. D.; Guastella, D.; Snyder, S. H. Bilirubin, formed by activation of heme oxygenase-2, protects neurons against oxidative stress injury. *Proc. Natl. Acad. Sci. USA* **96**:2445–2450; 1999.
- [57] Lleusy, S. F.; Tomaro, M. L. Evidence of involvement of bilirubin as physiological protector against oxidative damage. *Biochim. Biophys. Acta* **1223**:9–14; 1994.
- [58] Ossola, J. O.; Tomaro, M. L. Heme oxygenase induction by cadmium chloride: evidence for oxidative stress involvement. *Toxicology* **104**:141–147; 1995.
- [59] Ossola, J. O.; Tomaro, M. L. Heme oxygenase induction by UVA radiation. A response to oxidative stress in rat liver. *Int. J. Biochem. Cell Biol.* **30**:285–292; 1998.
- [60] Clark, J. E.; Foresti, R.; Green, C. J.; Motterlini, R. Dynamics of haem oxygenase-1 expression and bilirubin production in cellular protection against oxidative stress. *Biochem. J.* **348** (Pt 3):615–619; 2000.
- [61] Clark, J. E.; Foresti, R.; Sarathchandra, P.; Kaur, H.; Green, C. J.; Motterlini, R. Heme oxygenase-1-derived bilirubin ameliorates

- postischemic myocardial dysfunction. *Am. J. Physiol. Heart Circ. Physiol.* **278**:H643–H651; 2000.
- [62] Mazza, F.; Goodman, A. I.; Lombardo, G.; Vanella, A.; Abraham, N. G. Heme oxygenase I gene expression attenuates angiotensin II-mediated DNA damage in endothelial cells. *Exp. Biol. Med.* **228**: 576–583; 2003.
- [63] Morita, T.; Imai, T.; Yamaguchi, T.; Sugiyama, T.; Katayama, S.; Yoshino, G. Induction of heme oxygenase-1 in monocytes suppresses angiotensin II-elicited chemotactic activity through inhibition of CCR2: role of bilirubin and carbon monoxide generated by the enzyme. *Antioxid. Redox. Signal.* **5**:439–447; 2003.
- [64] Kwak, J. Y.; Takeshige, K.; Cheung, B. S.; Minakami, S. Bilirubin inhibits the activation of superoxide-producing NADPH oxidase in a neutrophil cell-free system. *Biochim. Biophys. Acta* **1076**:369–373; 1991.
- [65] Sano, K.; Nakamura, H.; Matsuo, T. Mode of inhibitory action of bilirubin on protein kinase C. *Pediatr. Res.* **19**:587–590; 1985.
- [66] Rajagopalan, S.; Kurz, S.; Munzel, T.; Tarpey, M.; Freeman, B. A.; Griending, K. K.; Harrison, D. G. Angiotensin II-mediated hypertension in the rat increases vascular superoxide production via membrane NADH/NADPH oxidase activation. Contribution to alterations of vasomotor tone. *J. Clin. Invest.* **97**:1916–1923; 1996.
- [67] Ishizaka, N.; Aizawa, T.; Mori, I.; Taguchi, J.; Yazaki, Y.; Nagai, R.; Ohno, M. Heme oxygenase-1 is upregulated in the rat heart in response to chronic administration of angiotensin II. *Am. J. Physiol. Heart Circ. Physiol.* **279**:H672–H678; 2000.
- [68] Sedlak, T. W.; Snyder, S. H. Bilirubin benefits: cellular protection by a biliverdin reductase antioxidant cycle. *Pediatrics* **113**:1776–1782; 2004.
- [69] Turkseven, S.; Kruger, A.; Mingone, C.J.; Kaminski, P.; Inaba, M.; Ikehara, S.; Wolin, M.S.; Abraham, N.G. The antioxidant mechanism of heme oxygenase-1 involves an increase in superoxide dismutase and catalase in experimental diabetes. *Am. J. Physiol.* In Press; 2005.
- [70] McClung, J. A.; Morita, T.; Rodella, L.; Rezzani, R.; Weiss, M. B.; Abraham, N. G. Heme oxygenase-1 prevents superoxide anion-associated vascular smooth muscle growth and decreases circulating endothelial cells in a rat model of balloon injury and restenosis in diabetes mellitus (abstract). *Circulation* **110**. American Heart Association Scientific Session; 2004.
- [71] Rezzani, R.; Quan, S.; Rodella, L.; Bianchi, R.; Goodman, A.; Abraham, N. G. Heme oxygenase-1 upregulation attenuates glucose-mediated oxidative stress renal injury in HO-2 knockout mice (abstract). *Hypertension* **42**; 2003.
- [72] Milstien, S.; Katusic, Z. Oxidation of tetrahydrobiopterin by peroxynitrite: implications for vascular endothelial function. *Biochem. Biophys. Res. Commun.* **263**:681–684; 1999.
- [73] Foresti, R.; Sarathchandra, P.; Clark, J. E.; Green, C. J.; Motterlini, R. Peroxynitrite induces haem oxygenase-1 in vascular endothelia to apoptosis. *Biochem. J.* **339**:729–736; 1999.
- [74] Chang, S. H.; Barbosa-Tessmann, I.; Chen, C.; Kilberg, M. S.; Agarwal, A. Glucose deprivation induces heme oxygenase-1 gene expression by a pathway independent of the unfolded protein response. *J. Biol. Chem.* **277**:1933–1940; 2002.
- [75] Mingone, C. J.; Turkseven, S.; Wolin, M. S.; Abraham, N. G. Heme-oxygenase-1 modulates vascular responses in diabetic rats via guanylate cyclase activation: role of superoxide dismutase (abstract). *FASEB*; 2005.
- [76] Mansoor, A.; Mingone, C. J.; Turkseven, S.; Wolin, M. S.; Abraham, N. G. Down regulation of iNOS and eNOS by overexpression of heme oxygenase-1 restore vascular response in diabetic rats (abstract). *FASEB*; 2005.
- [77] Hill-Kapturczak, N.; Chang, S. H.; Agarwal, A. Heme oxygenase and the kidney. *DNA Cell Biol.* **21**:307–321; 2002.
- [78] Vachharajani, T. J.; Work, J.; Issekutz, A. C.; Granger, D. N. Heme oxygenase modulates selectin expression in different regional vascular beds. *Am. J. Physiol. Heart Circ. Physiol.* **278**:H1613–H1617; 2000.
- [79] Wang, W. W.; Smith, D. L.; Zucker, S. D. Bilirubin inhibits iNOS expression and NO production in response to endotoxin in rats. *Hepatology* **40**:424–433; 2004.
- [80] Sacerdoti, D.; Escalante, B.; Abraham, N. G.; McGiff, J. C.; Levere, R. D.; Schwartzman, M. L. Treatment with tin prevents the development of hypertension in spontaneously hypertensive rats. *Science* **243**:388–390; 1989.
- [81] Solari, V.; Piotrowska, A. P.; Puri, P. Expression of heme oxygenase-1 and endothelial nitric oxide synthase in the lung of newborns with congenital diaphragmatic hernia and persistent pulmonary hypertension. *J. Pediatr. Surg.* **38**:808–813; 2003.
- [82] Snyder, S. H.; Baranano, D. E. Heme oxygenase: a font of multiple messengers. *Neuropsychopharmacology* **25**:294–298; 2001.
- [83] Motterlini, R.; Foresti, R.; Intaglietta, M.; Winslow, R. M. NO-mediated activation of heme oxygenase: endogenous cytoprotection against oxidative stress to endothelium. *Am. J. Physiol.* **270**: H107–H114; 1996.
- [84] Stanford, S. J.; Walters, M. J.; Hislop, A. A.; Haworth, S. G.; Evans, T. W.; Mann, B. E.; Motterlini, R.; Mitchell, J. A. Heme oxygenase is expressed in human pulmonary artery smooth muscle where carbon monoxide has an anti-proliferative role. *Eur. J. Pharmacol.* **473**:135–141; 2003.
- [85] Ndisang, J. F.; Tabien, H. E.; Wang, R. Carbon monoxide and hypertension. *J. Hypertens.* **22**:1057–1074; 2004.
- [86] Ndisang, J. F.; Wu, L.; Zhao, W.; Wang, R. Induction of heme oxygenase-1 and stimulation of cGMP production by hemin in aortic tissues from hypertensive rats. *Blood* **101**:3893–3900; 2003.
- [87] Ou, H. S.; Yang, J.; Dong, L. W.; Pang, Y. Z.; Su, J. Y.; Tang, C. S.; Liu, N. K. Role of endogenous carbon monoxide in the pathogenesis of hypotension during septic shock. *Sheng Li Xue. Bao.* **51**:1–6; 1999.
- [88] Beltowski, J.; Jamroz, A.; Borkowska, E. [Heme oxygenase and carbon monoxide in the physiology and pathology of the cardiovascular system]. *Postepy Hig. Med. Dosw. (Online)* **58**:83–99; 2004.
- [89] Ushiyama, M.; Morita, T.; Katayama, S. Carbon monoxide regulates blood pressure cooperatively with nitric oxide in hypertensive rats. *Heart Vessels* **16**:189–195; 2002.
- [90] Togane, Y.; Morita, T.; Suematsu, M.; Ishimura, Y.; Yamazaki, J. I.; Katayama, S. Protective roles of endogenous carbon monoxide in neointimal development elicited by arterial injury. *Am. J. Physiol. Heart Circ. Physiol.* **278**:H623–H632; 2000.
- [91] Otterbein, L. E.; Zuckerbraun, B. S.; Haga, M.; Liu, F.; Song, R.; Usheva, A.; Stachulak, C.; Bodyak, N.; Smith, R. N.; Cszizmadia, E.; Tyagi, S.; Akamatsu, Y.; Flavell, R. J.; Billiar, T. R.; Tzeng, E.; Bach, F. H.; Choi, A. M.; Soares, M. P. Carbon monoxide suppresses arteriosclerotic lesions associated with chronic graft rejection and with balloon injury. *Nat. Med.* **9**:183–190; 2003.
- [92] Kukoba, T. V.; Moibenko, O. O.; Kotsioruba, A. V. [Cardio-protective effect of heme oxygenase-1 induction by hemin on the isolated rat heart during ischemia–reperfusion]. *Fiziol. Zh.* **49**:14–21; 2003.
- [93] Yamada, N.; Yamaya, M.; Okinaga, S.; Nakayama, K.; Sekizawa, K.; Shibahara, S.; Sasaki, H. Microsatellite polymorphism in the heme oxygenase-1 gene promoter is associated with susceptibility to emphysema. *Am. J. Hum. Genet.* **66**:187–195; 2000.
- [94] Tulis, D. A.; Durante, W.; Liu, X.; Evans, A. J.; Peyton, K. J.; Schafer, A. I. Adenovirus-mediated heme oxygenase-1 gene delivery inhibits injury-induced vascular neointima formation. *Circulation* **104**:2710–2715; 2001.
- [95] Duckers, H. J.; Boehm, M.; True, A. L.; Yet, S. F.; San, H.; Park, J. L.; Clinton, W. R.; Lee, M. E.; Nabel, G. J.; Nabel, E. G. Heme oxygenase-1 protects against vascular constriction and proliferation. *Nat. Med.* **7**:693–698; 2001.

- [96] Li Volti, G.; Wang, J.; Traganos, F.; Kappas, A.; Abraham, N. G. Differential effect of heme oxygenase-1 in endothelial and smooth muscle cell cycle progression. *Biochem. Biophys. Res. Commun.* **296**:1077–1082; 2002.
- [97] Kushida, T.; Quan, S.; Yang, L.; Ikehara, S.; Kappas, A.; Abraham, N. G. A significant role for the heme oxygenase-1 gene in endothelial cell cycle progression. *Biochem. Biophys. Res. Commun.* **291**:68–75; 2002.
- [98] Agarwal, A.; Balla, J.; Alam, J.; Croatt, A. J.; Nath, K. A. Induction of heme oxygenase in toxic renal injury: a protective role in cisplatin nephrotoxicity in the rat. *Kidney Int.* **48**:1298–1307; 1995.
- [99] Lutton, J. D.; da Silva, J.-L.; Moqattash, S.; Brown, A. C.; Levere, R. D.; Abraham, N. G. Differential induction of heme oxygenase in the hepatocarcinoma cell line (Hep3b) by environmental agents. *J. Cell. Biochem.* **49**:259–265; 1992.
- [100] Gong, J.; Traganos, F. D. Z. Growth imbalance and altered expression of cyclins B1, A, E and D3 in MOLT-4 cells synchronized in the cell cycle by inhibitors of DNA replication. *Cell Growth Differ.* **6**:1485–1493; 1995.
- [101] Mantovani, A.; Bussofino, F.; Introna, M. Cytokine regulation of endothelial cell function: from molecular level to the bedside. *Immunol. Today* **18**:231–240; 1997.
- [102] Abraham, N. G.; Scapagnini, G.; Kappas, A. Human heme oxygenase: cell cycle-dependent expression and DNA microarray identification of multiple gene responses after transduction of endothelial cells. *J. Cell. Biochem.* **90**:1098–1111; 2003.
- [103] Kong, D.; Melo, L. G.; Mangi, A. A.; Zhang, L.; Lopez-Illasaca, M.; Perrella, M. A.; Liew, C. C.; Pratt, R. E.; Dzau, V. J. Enhanced inhibition of neointimal hyperplasia by genetically engineered endothelial progenitor cells. *Circulation* **109**:1769–1775; 2004.
- [104] Kanai, M.; Akaba, K.; Sasaki, A.; Sato, M.; Harano, T.; Shibahara, S.; Kurachi, H.; Yoshida, T.; Hayasaka, K. Neonatal hyperbilirubinemia in Japanese neonates: analysis of the heme oxygenase-1 gene and fetal hemoglobin composition in cord blood. *Pediatr. Res.* **54**:165–171; 2003.
- [105] Ono, K.; Mannami, T.; Iwai, N. Association of a promoter variant of the haeme oxygenase-1 gene with hypertension in women. *J. Hypertens.* **21**:1497–1503; 2003.
- [106] Kanai, M.; Tanabe, S.; Okada, M.; Suzuki, H.; Niki, T.; Katsuura, M.; Akiba, T.; Hayasaka, K. Polymorphisms of heme oxygenase-1 and bilirubin UDP-glucuronosyltransferase genes are not associated with Kawasaki disease susceptibility. *J. Tohoku, Exp. Med.* **200**:155–159; 2003.
- [107] Chang, K. W.; Lee, T. C.; Yeh, W. I.; Chung, M. Y.; Liu, C. J.; Chi, L. Y.; Lin, S. C. Polymorphism in heme oxygenase-1 (HO-1) promoter is related to the risk of oral squamous cell carcinoma occurring on male areca chewers. *Br. J. Cancer* **91**:1551–1555; 2004.
- [108] Funk, M.; Endler, G.; Schillinger, M.; Mustafa, S.; Hsieh, K.; Exner, M.; Lalouschek, W.; Mannhalter, C.; Wagner, O. The effect of a promoter polymorphism in the heme oxygenase-1 gene on the risk of ischaemic cerebrovascular events: the influence of other vascular risk factors. *Thromb. Res.* **113**:217–223; 2004.
- [109] Schillinger, M.; Exner, M.; Minar, E.; Mlekusch, W.; Mullner, M.; Mannhalter, C.; Bach, F. H.; Wagner, O. Heme oxygenase-1 genotype and restenosis after balloon angioplasty: a novel vascular protective factor. *J. Am. Coll. Cardiol.* **43**:950–957; 2004.
- [110] Gu, Y.; Ortines, R. Heme oxygenase-1 is cardioprotective in the remodeled, failing heart (abstract). *The American Heart Association Scientific Sessions 2004*; 2004.
- [111] Seki, T.; Naruse, M.; Naruse, K.; Yoshimoto, T.; Tanabe, A.; Seki, M.; Tago, K.; Imaki, T.; Demura, R.; Demura, H. Induction of heme oxygenase produces load-independent cardioprotective effects in hypertensive rats. *Life Sci.* **65**:1077–1086; 1999.
- [112] Ooi, H.; Ito, M.; Pimental, D. Heme oxygenase protects against  $\beta$ -adrenergic receptor-stimulated apoptosis in cardiac (abstract). *The American Heart Association Scientific Sessions 2004*; 2004.
- [113] Kawamoto, S.; Flynn, J. P.; Allen, M. D. Heme oxygenase-1 induction improves the viability of adult cardiomyocyte cellular grafts in vivo (abstract). *The American Heart Association Scientific Sessions 2004*; 2004.
- [114] Tang, Y. L.; Tang, Y.; Zhang, Y. C.; Philips, M. I. Mesenchymal stem cells protected with a hypoxia-regulated heme oxygenase-1 vector: A potential strategy to prevent graft cell death in the ischemic heart (abstract). *Circulation* **110**:17; 2004.
- [115] Motterlini, R.; Clark, J. E.; Foresti, R.; Sarathchandra, P.; Mann, B. E.; Green, C. J. Carbon monoxide-releasing molecules: characterization of biochemical and vascular activities. *Circ. Res.* **90**:E17–E24; 2002.
- [116] Chatterjee, P. K. Water-soluble carbon monoxide-releasing molecules: helping to elucidate the vascular activity of the 'silent killer'. *Br. J. Pharmacol.* **142**:391–393; 2004.
- [117] Clark, J. E.; Naughton, P.; Shurey, S.; Green, C. J.; Johnson, T. R.; Mann, B. E.; Foresti, R.; Motterlini, R. Cardioprotective actions by a water-soluble carbon monoxide-releasing molecule. *Circ. Res.* **93**:000; 2003.
- [118] Foresti, R.; Hammad, J.; Clark, J. E.; Johnson, T. R.; Mann, B. E.; Friebe, A.; Green, C. J.; Motterlini, R. Vasoactive properties of CORM-3, a novel water-soluble carbon monoxide-releasing molecule. *Br. J. Pharmacol.* **142**:453–460; 2004.
- [119] Wang, R.; Wang, Z.; Wu, L. Carbon monoxide-induced vasorelaxation and the underlying mechanisms. *Br. J. Pharmacol.* **121**:927–934; 1997.
- [120] Morita, T.; Perrella, M. A.; Lee, M.; Kourembanas, S. Smooth muscle cell-derived carbon monoxide is a regulator of vascular cGMP. *Proc. Natl. Acad. Sci. USA* **92**:1475–1479; 1995.
- [121] Ndisang, J. F.; Wang, R. Alterations in heme oxygenase/carbon monoxide system in pulmonary arteries in hypertension. *Exp. Biol. Med. (Maywood)* **228**:557–563; 2003.
- [122] Levere, R. D.; Martasek, P.; Escalante, B.; Schwartzman, M. L.; Abraham, N. G. Effect of heme arginate administration on blood pressure in spontaneously hypertensive rats. *J. Clin. Invest.* **86**:213–219; 1990.
- [123] Sammut, I. A.; Foresti, R.; Clark, J. E.; Exon, D. J.; Vesely, M. J. J.; Sarathchandra, P.; Green, C. J.; Motterlini, R. Carbon monoxide is a major contributor to the regulation of vascular tone in aortas expressing high levels of haeme oxygenase-1. *Br. J. Pharmacol.* **125**:1437–1444; 1998.
- [124] Motterlini, R.; Mann, B. E.; Johnson, T. R.; Clark, J. E.; Foresti, R.; Green, C. J. Bioactivity and pharmacological actions of carbon monoxide-releasing molecules. *Curr. Pharm. Des.* **9**:2525–2539; 2003.
- [125] Abraham, N. G.; Mieval, P. A.; Quan, S.; Yang, L.; Burke-Wolin, T.; Mingone, C. J.; Goodman, A. I.; Nasjletti, A.; Wolin, M. S. Modulation of cyclic GMP by retrovirus-mediated human heme oxygenase-1 gene transfer in microvessel endothelial cells. *Am. J. Physiol.* **283**:L1117–L1124; 2002.
- [126] da-Silva, J. L.; Morishita, T.; Escalante, B.; Staudinger, R.; Drummond, G.; Goligorsky, M. S.; Lutton, J. D.; Abraham, N. G. Dual role of heme oxygenase in epithelial cell injury: contrasting effects of short-term and long-term exposure to oxidant stress. *J. Lab. Clin. Med.* **128**:290–296; 1996.
- [127] Abraham, N. G.; Lin, J. H.; Schwartzman, M. L.; Levere, R. D.; Shibahara, S. The physiological significance of heme oxygenase. *Int. J. Biochem.* **20**:543–558; 1988.
- [128] Kierner, A. K.; Bildner, N.; Weber, N. C.; Vollmar, A. M. Characterization of heme oxygenase 1 (heat shock protein 32) induction by atrial natriuretic peptide in human endothelial cells. *Endocrinology* **144**:802–812; 2003.
- [129] Polte, T.; Hemmerle, A.; Berndt, G.; Grosser, N.; Abate, A.; Schroder, H. Atrial natriuretic peptide reduces cyclosporin toxicity

- in renal cells: role of cGMP and heme oxygenase-1. *Free Radic. Biol. Med.* **32**:56–63; 2002.
- [130] Visner, G. A.; Lu, F.; Zhou, H.; Liu, J.; Kazemfar, K.; Agarwal, A. Rapamycin induces heme oxygenase-1 in human pulmonary vascular cells: implications in the antiproliferative response to rapamycin. *Circulation* **107**:911–916; 2003.
- [131] Polte, T.; Abate, A.; Dennery, P. A.; Schroder, H. Heme oxygenase-1 is a cGMP-inducible endothelial protein and mediates the cytoprotective action of nitric oxide. *Arterioscler. Thromb. Vasc. Biol.* **20**:1209–1215; 2000.
- [132] Kozma, F.; Johnson, R. A.; Zhang, F.; Yu, C.; Tong, X.; Nasjletti, A. Contribution of endogenous carbon monoxide to regulation of diameter in resistance vessels. *Am. J. Physiol.* **276**:R1087–R1094; 1999.
- [133] Wang, T.; Sterling, H.; Shao, W. A.; Yan, Q.; Bailey, M. A.; Giebisch, G.; Wang, W. H. Inhibition of heme oxygenase decreases sodium and fluid absorption in the loop of Henle. *Am. J. Physiol. Renal Physiol.* **285**:F484–F490; 2003.
- [134] Lim, I.; Gibbons, S. J.; Lyford, G. L.; Miller, S. M.; Strega, P. R.; Sarr, M. G.; Chatterjee, S.; Szurszewski, J. H.; Shah, V. H.; Farrugia, G. Carbon monoxide activates human intestinal smooth muscle L-type Ca<sup>2+</sup> channels through a nitric oxide-dependent mechanism. *Am. J. Physiol. Gastrointest. Liver Physiol.*; 2004.
- [135] Smallegange, C.; Hale, T. M.; Bushfield, T. L.; Adams, M. A. Persistent lowering of pressure by transplanting kidneys from adult spontaneously hypertensive rats treated with brief antihypertensive therapy. *Hypertension* **44**:89–94; 2004.
- [136] Nath, K. A.; Balla, J.; Jacob, H. S.; Vercellotti, G. M.; Levitt, M.; Rosenberg, M. E. Induction of heme oxygenase is a rapid protective response in rhabdomyolysis in the rat. *J. Clin. Invest.* **90**:267–270; 1992.
- [137] Mosley, K.; Wembridge, D. E.; Cattell, V.; Cook, H. T. Heme oxygenase is induced in nephrotoxic nephritis and hemin, a stimulator of heme oxygenase synthesis, ameliorates disease. *Kidney Int.* **53**:672–678; 1998.
- [138] Arregui, B.; Lopez, B.; Garcia, S. M.; Valero, F.; Navarro, C.; Fenoy, F. J. Acute renal hemodynamic effects of dimanganese decacarbonyl and cobalt protoporphyrin. *Kidney Int.* **65**:564–574; 2004.
- [139] Mustafa, M. R.; Johns, E. J. The role of haem oxygenase in renal vascular reactivity in normotensive and hypertensive rats. *J. Hypertens.* **19**:1105–1111; 2001.
- [140] Rodriguez, F.; Kemp, R.; Balazy, M.; Nasjletti, A. Effects of exogenous heme on renal function. Role of heme oxygenase and cyclooxygenase. *Hypertension* **42**:680–684; 2003.
- [141] Raizada, M. K.; Francis, S. C.; Wang, H.; Gelband, C. H.; Reaves, P. Y.; Katovich, M. J. Targeting of the renin-angiotensin system by antisense gene therapy: a possible strategy for the long-term control of hypertension. *J. Hypertens.* **18**:353–362; 2000.
- [142] Laursen, J. B.; Rajagopalan, S.; Galis, Z.; Tarpey, M.; Freeman, B. A.; Harrison, D. G. Role of superoxide in angiotensin II-induced but not catecholamine-induced hypertension. *Circulation* **95**:588–593; 1997.
- [143] Haugen, E. N.; Croatt, A. J.; Nath, K. A. Angiotensin II induces renal oxidant stress in vivo and heme oxygenase-1 in vivo and in vitro. *Kidney Int.* **58**:144–152; 2000.
- [144] Aizawa, T.; Ishizaka, N.; Taguchi, J.; Nagai, R.; Mori, I.; Tang, S. S.; Ingelfinger, J. R.; Ohno, M. Heme oxygenase-1 is upregulated in the kidney of angiotensin II-induced hypertensive rats: possible role in renoprotection. *Hypertension* **35**:800–806; 2000.
- [145] Ortiz, P. A.; Garvin, J. L. Superoxide stimulates NaCl absorption by the thick ascending limb. *Am. J. Physiol. Renal Physiol.* **283**:F957–F962; 2002.
- [146] Datta, P. K.; Moulder, J. E.; Fish, B. L.; Cohen, E. P.; Lianos, E. A. Induction of heme oxygenase 1 in radiation nephropathy: role of angiotensin II. *Radiat. Res.* **155**:734–739; 2001.
- [147] Ishizaka, N.; Griendling, K. K. Heme oxygenase-1 is regulated by angiotensin II in rat vascular smooth muscle cells. *Hypertension* **29**:790–795; 1997.
- [148] Poss, K. D.; Tonegawa, S. Heme oxygenase 1 is required for mammalian iron reutilization. *Proc. Natl. Acad. Sci. USA* **94**:10919–10924; 1997.
- [149] Yachie, A.; Niida, Y.; Wada, T.; Igarashi, N.; Kaneda, H.; Toma, T.; Ohta, K.; Kasahara, Y.; Koizumi, S. Oxidative stress causes enhanced endothelial cell injury in human heme oxygenase-1 deficiency. *J. Clin. Invest.* **103**:129–135; 1999.
- [150] Ishizaka, N.; De Leon, H.; Laursen, J. B.; Fukui, T.; Wilcox, J. N.; De Keulenaer, G.; Griendling, K. K.; Alexander, R. W. Angiotensin II-induced hypertension increases heme oxygenase-1 expression in rat aorta. *Circulation* **96**:1923–1929; 1997.
- [151] Fukui, T.; Ishizaka, N.; Rajagopalan, S.; Laursen, J. B.; Capers, Q.; Taylor, W. R.; Harrison, D. G.; De Leon, H.; Wilcox, J. N.; Griendling, K. K. p22phox mRNA expression and NADPH oxidase activity are increased in aortas from hypertensive rats. *Circ. Res.* **80**:45–51; 1997.
- [152] Sun, C.; Sellers, K. W.; Summers, C.; Raizada, M. K. NAD(P)H oxidase inhibition attenuates neuronal chronotropic actions of angiotensin II. *Circ. Res.*; 2005.
- [153] Wolf, G. Free radical production and angiotensin. *Curr. Hypertens. Rep.* **2**:167–173; 2000.
- [154] Botros, F. T.; Laniado-Schwartzman, M.; Abraham, N. G. Regulation of cyclooxygenase- and cytochrome p450-derived eicosanoids by heme oxygenase in the rat kidney. *Hypertension* **39**:639–644; 2002.
- [155] Li Volti, G.; Seta, F.; Schwartzman, M. L.; Nasjletti, A.; Abraham, N. G. Heme oxygenase attenuates angiotensin II-mediated increase in cyclooxygenase-2 activity in human femoral endothelial cells. *Hypertension* **41**:715–719; 2003.
- [156] Escalante, B.; Sessa, W. C.; Falck, J. R.; Yadagiri, P.; Schwartzman, M. L. Vasoactivity of 20-hydroxyeicosatetraenoic acid is dependent on metabolism by cyclooxygenase. *J. Pharmacol. Exp. Ther.* **248**:229–232; 1988.
- [157] Zou, A. P.; Ma, Y. H.; Sui, Z. H.; Ortiz-de-Montellano, P. R.; Clark, J. E.; Masters, B. S.; Roman, R. J. Effects of 17-octadecynoic acid, a suicide-substrate inhibitor of cytochrome P450 fatty acid omega-hydroxylase, on renal function in rats. *J. Pharmacol. Exp. Ther.* **268**:474–481; 1994.
- [158] Wilcox, C. S.; Welch, W. J. Thromboxane mediation of the pressor response to infused angiotensin II. *Am. J. Hypertens.* **3**:242–249; 1990.
- [159] Lin, L.; Balazy, M.; Pagano, P. J.; Nasjletti, A. Expression of prostaglandin H<sub>2</sub>-mediated mechanism of vascular contraction in hypertensive rats. Relation to lipoxygenase and prostacyclin synthase activities. *Circ. Res.* **74**:197–205; 1994.
- [160] Sessa, W. C.; Abraham, N. G.; Escalante, B.; Schwartzman, M. L. Manipulation of cytochrome P-450 dependent renal thromboxane synthase activity in spontaneously hypertensive rats. *J. Hypertens.* **7**:37–42; 1989.
- [161] Haider, A.; Olszanecki, R.; Gryglewski, R.; Schwartzman, M. L.; Lianos, E.; Kappas, A.; Nasjletti, A.; Abraham, N. G. Regulation of cyclooxygenase by the heme-heme oxygenase system in microvessel endothelial cells. *J. Pharmacol. Exp. Ther.* **300**:188–194; 2002.
- [162] da-Silva, J. L.; Tiefenthaler, M.; Park, E.; Escalante, B.; Schwartzman, M. L.; Levere, R. D.; Abraham, N. G. Tin-mediated heme oxygenase gene activation and cytochrome P450 arachidonate hydroxylase inhibition in spontaneously hypertensive rats [published erratum appears in *Am. J. Med. Sci.* 308(2):138; 1994]. *Am. J. Med. Sci.* **307**:173–181; 1994.
- [163] Sacerdoti, D.; Abraham, N. G.; McGiff, J. C.; Schwartzman, M. L. Renal cytochrome P450-dependent metabolism of arachidonic acid in spontaneously hypertensive rats. *Biochem. Pharmacol.* **37**:521–527; 1988.

- [164] McGiff, J. C. Cytochrome P-450 metabolism of arachidonic acid. *Annu. Rev. Pharmacol. Toxicol.* **31**:339–369; 1991.
- [165] McGiff, J. C. Eicosanoids and hypertension. *Rev. Port. Cardiol.* **7**:59–61; 1988.
- [166] Schwartzman, M. L.; Falck, J. R.; Yadagiri, P.; Escalante, B. Metabolism of 20-hydroxyeicosatetraenoic acid by cyclooxygenase. Formation and identification of novel endothelium-dependent vasoconstrictor metabolites. *J. Biol. Chem.* **264**:11658–11662; 1989.
- [167] Carroll, M. A.; Garcia, M. P.; Falck, J. R.; McGiff, J. C. Cyclooxygenase dependency of the renovascular actions of cytochrome P450-derived arachidonate metabolites. *J. Pharmacol. Exp. Ther.* **260**:104–109; 1992.
- [168] Ma, Y. H.; Gebremedhin, D.; Schwartzman, M. L.; Falck, J. R.; Clark, J. E.; Masters, B. S.; Harder, D. R.; Roman, R. J. 20-Hydroxyeicosatetraenoic acid is an endogenous vasoconstrictor of canine renal arcuate arteries. *Circ. Res.* **72**:126–136; 1993.
- [169] Schwartzman, M.; Ferreri, N. R.; Carroll, M. A.; Songu-Mize, E.; McGiff, J. C. Renal cytochrome P450-related arachidonate metabolite inhibits (Na<sup>+</sup> + K<sup>+</sup>)ATPase. *Nature* **314**:620–622; 1985.
- [170] Zou, A. P.; Fleming, J. T.; Falck, J. R.; Jacobs, E. R.; Gebremedhin, D.; Harder, D. R.; Roman, R. J. 20-HETE is an endogenous inhibitor of the large-conductance Ca(2+)-activated K<sup>+</sup> channel in renal arterioles. *Am. J. Physiol.* **270**:R228–R237; 1996.
- [171] Zou, A. P.; Drummond, H. A.; Roman, R. J. Role of 20-HETE in elevating loop chloride reabsorption in Dahl SS/Jr rats. *Hypertension* **27**:631–635; 1996.
- [172] Wang, W.; Lu, M. Effect of arachidonic acid on activity of the apical K<sup>+</sup> channel in the thick ascending limb of the rat kidney. *J. Gen. Physiol.* **106**:727–743; 1995.
- [173] Zou, A. P.; Imig, J. D.; Ortiz-de-Montellano, P. R.; Sui, Z.; Falck, J. R.; Roman, R. J. Effect of P-450 omega-hydroxylase metabolites of arachidonic acid on tubuloglomerular feedback. *Am. J. Physiol.* **266**:F934–F941; 1994.
- [174] Lin, F.; Rios, A.; Falck, J. R.; Belosludtsev, Y.; Schwartzman, M. L. 20-Hydroxyeicosatetraenoic acid is formed in response to EGF and is a mitogen in rat proximal tubule. *Am. J. Physiol.* **269**:F806–F816; 1995.
- [175] Roman, R. J. P-450 metabolites of arachidonic acid in the control of cardiovascular function. *Physiol. Rev.* **82**:131–185; 2002.
- [176] Sabaawy, H. E.; Zhang, F.; Nguyen, X.; Elhosseiny, A.; Nasjletti, A.; Schwartzman, M.; Dennerly, P.; Kappas, A.; Abraham, N. G. Human heme oxygenase-1 gene transfer lowers blood pressure and promotes growth in spontaneously hypertensive rats. *Hypertension* **38**:210–215; 2001.
- [177] Martasek, P.; Schwartzman, M. L.; Goodman, A. I.; Solangi, K. B.; Levere, R. D.; Abraham, N. G. Hemin and L-arginine regulation of blood pressure in spontaneous hypertensive rats. *J. Am. Soc. Nephrol.* **2**:1078–1084; 1991.
- [178] Chemick, R. J.; Martasek, P.; Levere, R. D.; Margreiter, R.; Abraham, N. G. Sensitivity of human tissue heme oxygenase to a new synthetic metalloporphyrin. *Hepatology* **10**:365–369; 1989.
- [179] Rodriguez, F.; Lamon, B. D.; Gong, W.; Kemp, R.; Nasjletti, A. Nitric oxide synthesis inhibition promotes renal production of carbon monoxide. *Hypertension* **43**:347–351; 2004.
- [180] Johnson, R. A.; Lavesa, M.; DeSeyn, K.; Scholer, M. J.; Nasjletti, A. Heme oxygenase substrates acutely lower blood pressure in hypertensive rats. *Am. J. Physiol.* **271**:H1132–H1138; 1996.
- [181] da Silva, J. L.; Zand, B. A.; Yang, L. M.; Sabaawy, H. E.; Lianos, E.; Abraham, N. G. Heme oxygenase isoform-specific expression and distribution in the rat kidney. *Kidney Int.* **59**:1448–1457; 2001.
- [182] Botros, F. T.; Schwartzman, M. L.; Abraham, N. G. Regulation of cyclooxygenase- and cytochrome P450-derived eicosanoids by heme oxygenase isoforms in rat kidney (abstract). *Hypertension* **38** (3); 2001.
- [183] Abraham, N. G. Heme oxygenase attenuated angiotensin II-mediated increase in cyclooxygenase activity and decreased isoprostane F2alpha in endothelial cells. *Thromb. Res.* **110**:305–309; 2003.
- [184] Zhang, F.; Kaide, J. I.; Rodriguez-Mulero, F.; Abraham, N. G.; Nasjletti, A. Vasoregulatory function of the heme-heme oxygenase-carbon monoxide system. *Am. J. Hypertens.* **14**:62S–67S; 2001.
- [185] Goodman, A. I.; Quan, S.; Yang, L.; Synghal, A.; Abraham, N. G. Functional expression of human heme oxygenase-1 gene in renal structure of spontaneously hypertensive rats. *Exp. Biol. Med. (Maywood)* **228**:454–458; 2003.
- [186] Smith, W. L.; Bell, T. G. Immunohistochemical localization of the prostaglandin-forming cyclooxygenase in renal cortex. *Am. J. Physiol.* **235**:F451–F457; 1978.
- [187] Harris, R. C.; McKanna, J. A.; Akai, Y.; Jacobson, H. R.; DuBois, R. N.; Breyer, M. D. Cyclooxygenase-2 is associated with the macula densa of rat kidney and increases with salt restriction. *J. Clin. Invest.* **94**:2504–2510; 1994.
- [188] Vio, C. P.; Cespedes, C.; Gallardo, P.; Masferrer, J. L. Renal identification of cyclooxygenase-2 in a subset of thick ascending limb cells. *Hypertension* **30**:687–692; 1997.
- [189] Traynor, T. R.; Smart, A.; Briggs, J. P.; Schnermann, J. Inhibition of macula densa-stimulated renin secretion by pharmacological blockade of cyclooxygenase-2. *Am. J. Physiol.* **277**:F706–F710; 1999.
- [190] Carmines, P. K.; Bell, P. D.; Roman, R. J.; Work, J.; Navar, L. G. Prostaglandins in the sodium excretory response to altered renal arterial pressure in dogs. *Am. J. Physiol.* **248**:F8–F14; 1985.
- [191] Yang, T.; Endo, Y.; Huang, Y. G.; Smart, A.; Briggs, J. P.; Schnermann, J. Renin expression in COX-2-knockout mice on normal or low-salt diets. *Am. J. Physiol. Renal Physiol.* **279**:F819–F825; 2000.
- [192] Cheng, H. F.; Wang, J. L.; Zhang, M. Z.; Wang, S. W.; McKanna, J. A.; Harris, R. C. Genetic deletion of COX-2 prevents increased renin expression in response to ACE inhibition. *Am. J. Physiol. Renal Physiol.* **280**:F449–F456; 2001.
- [193] Haider, A.; Olszanecki, R.; Gryglewski, R.; Schwartzman, M. L.; Lianos, E.; Kappas, A.; Nasjletti, A.; Abraham, N. G. Regulation of cyclooxygenase by the heme-heme oxygenase system in microvessel endothelial cells. *J. Pharmacol. Exp. Ther.* **300**:188–194; 2002.
- [194] Grana, X.; Reddy, E. P. Cell cycle control in mammalian cells: role of cyclins, cyclin dependent kinases (CDKs), growth suppressor genes and cyclin-dependent kinase inhibitors (CKIs). *Oncogene* **11**:211–219; 1995.
- [195] Peter, M.; Herskowitz, I. Joining the complex: cyclin-dependent kinase inhibitory proteins and the cell cycle. *Cell* **79**:181–184; 1994.
- [196] Polyak, K.; Kato, J. Y.; Solomon, M. J.; Sherr, C. J.; Massague, J.; Roberts, J. M.; Koff, A. p27Kip1, a cyclin-Cdk inhibitor, links transforming growth factor-beta and contact inhibition to cell cycle arrest. *Genes Dev.* **8**:9–22; 1994.
- [197] Juan, G.; Ardel, B.; Li, X.; Mikulski, S. M.; Shogen, K.; Ardel, W.; Mittelman, A.; Darzynkiewicz, Z. G1 arrest of U937 cells by oncosis is associated with suppression of cyclin D3 expression, induction of p16INK4A, p21WAF1/CIP1 and p27KIP and decreased pRb phosphorylation. *Leukemia* **12**:1241–1248; 1998.
- [198] Darzynkiewicz, Z.; Gong, J.; Juan, G.; Ardel, B.; Traganos, F. Cytometry of cyclin proteins. *Cytometry* **25**:1–13; 1996.
- [199] Nakao-Hayashi, J.; Ito, H.; Kawashima, S. An oxidative mechanism is involved in high glucose-induced serum protein modification causing inhibition of endothelial cell proliferation. *Atherosclerosis* **97**:89–95; 1992.
- [200] Brouard, S.; Otterbein, L. E.; Anrather, J.; Tobiasch, E.; Bach, F. H.; Choi, A. M.; Soares, M. P. Carbon monoxide generated by heme



- oxygenase 1 suppresses endothelial cell apoptosis. *J. Exp. Med.* **192**:1015–1026; 2000.
- [201] Zhang, X.; Shan, P.; Otterbein, L. E.; Alam, J.; Flavell, R. A.; Davis, R. J.; Choi, A. M.; Lee, P. J. Carbon monoxide inhibition of apoptosis during ischemia-reperfusion lung injury is dependent on the p38 mitogen-activated protein kinase pathway and involves caspase 3. *J. Biol. Chem.* **278**:1248–1258; 2003.
- [202] Petrache, I.; Otterbein, L. E.; Alam, J.; Wiegand, G. W.; Choi, A. M. Heme oxygenase-1 inhibits TNF-alpha-induced apoptosis in cultured fibroblasts. *Am. J. Physiol. Lung Cell Mol. Physiol.* **278**:L312–L319; 2000.
- [203] Colombrina, C.; Lombardo, G.; Scapagnini, G.; Abraham, N. G. Heme oxygenase-1 expression levels are cell cycle dependent. *Biochem. Biophys. Res. Commun.* **308**:1001–1008; 2003.
- [204] Griendling, K. K.; Sorescu, D.; Ushio-Fukai, M. NAD(P)H oxidase: role in cardiovascular biology and disease. *Circ. Res.* **86**:494–501; 2000.
- [205] Katusic, Z. S. Superoxide anion and endothelial regulation of arterial tone. *Free Radic. Biol. Med.* **20**:443–448; 1996.
- [206] Koya, D.; Hayashi, K.; Kitada, M.; Kashiwagi, A.; Kikkawa, R.; Haneda, M. Effects of antioxidants in diabetes-induced oxidative stress in the glomeruli of diabetic rats. *J. Am. Soc. Nephrol.* **14**:S250–S253; 2003.
- [207] Quan, S.; Kaminski, P. M.; Yang, L.; Morita, T.; Inaba, M.; Ikehara, S.; Goodman, A. I.; Wolin, M. S.; Abraham, N. G. Heme oxygenase-1 prevents superoxide anion associated endothelial cell sloughing in diabetic rats. *Biochem. Biophys. Res. Commun.* **315**:509–516; 2004.
- [208] Grosser, N.; Abate, A.; Oberle, S.; Vreman, H. J.; Dennery, P. A.; Becker, J. C.; Pohle, T.; Seidman, D. S.; Schroder, H. Heme oxygenase-1 induction may explain the antioxidant profile of aspirin. *Biochem. Biophys. Res. Commun.* **308**:956–960; 2003.
- [209] Oberle, S.; Abate, A.; Grosser, N.; Hemmerle, A.; Vreman, H. J.; Dennery, P. A.; Schneider, H. T.; Stalleicken, D.; Schroder, H. Endothelial protection by pentaerythrityl trinitrate: bilirubin and carbon monoxide as possible mediators. *Exp. Biol. Med. (Maywood)* **228**:529–534; 2003.
- [210] Polte, T.; Oberle, S.; Schroder, H. The nitric oxide donor SIN-1 protects endothelial cells from tumor necrosis factor-alpha-mediated cytotoxicity: possible role for cyclic GMP and heme oxygenase. *J. Mol. Cell. Cardiol.* **29**:3305–3310; 1997.
- [211] Grosser, N.; Hemmerle, A.; Berndt, G.; Erdmann, K.; Hinkelmann, U.; Schurgerc, S.; Wijayanti, N.; Immenschuh, S.; Schroder, H. The antioxidant defense protein heme oxygenase 1 is a novel target for statins in endothelial cells. *Free Radic. Biol. Med.* **37**:2064–2071; 2004.
- [212] Lee, T. S.; Chang, C. C.; Zhu, Y.; Shyy, J. Y. Simvastatin induces heme oxygenase-1: a novel mechanism of vessel protection. *Circulation* **110**:1296–1302; 2004.
- [213] Grosser, N.; Erdmann, K.; Hemmerle, A.; Berndt, G.; Hinkelmann, U.; Smith, G.; Schroder, H. Rosuvastatin upregulates the antioxidant defense protein heme oxygenase-1. *Biochem. Biophys. Res. Commun.* **325**:871–876; 2004.
- [214] Abraham, N. G.; daSilva, J. L.; Dunn, M. W.; Kigasawa, K.; Shibahara, S. Retinal pigment epithelial cell-based gene therapy against hemoglobin toxicity. *Inter. J. Mol. Med.* **1**:657–663; 1998.
- [215] Stec, D. E. Smart gene therapy for the heart. *Hypertension* **43**:720–721; 2004.
- [216] Pachori, A. S.; Melo, L. G.; Hart, M. L.; Noiseux, N.; Zhang, L.; Morello, F.; Solomon, S. D.; Stahl, G. L.; Pratt, R. E.; Dzau, V. J. Hypoxia-regulated therapeutic gene as a preemptive treatment strategy against ischemia/reperfusion tissue injury. *Proc. Natl. Acad. Sci. USA* **101**:12282–12287; 2004.
- [217] Coito, A. J.; Buelow, R.; Shen, X. D.; Amersi, F.; Moore, C.; Volk, H. D.; Busuttil, R. W.; Kupiec-Weglinski, J. W. Heme oxygenase-1 gene transfer inhibits inducible nitric oxide synthase expression and protects genetically fat Zucker rat livers from ischemia-reperfusion injury. *Transplantation* **74**:96–102; 2002.
- [218] Vassalli, G.; Fleury, S.; Li, J.; Goy, J. J.; Kappenberger, L.; von Segesser, L. K. Gene transfer of cytoprotective and immunomodulatory molecules for prevention of cardiac allograft rejection. *Eur. J. Cardiothorac. Surg.* **24**:794–806; 2003.
- [219] Braudeau, C.; Bouchet, D.; Tesson, L.; Iyer, S.; Remy, S.; Buelow, R.; Anegon, I.; Chauveau, C. Induction of long-term cardiac allograft survival by heme oxygenase-1 gene transfer. *Gene Ther.* **11**:701–710; 2004.
- [220] Tang, Y. L.; Tang, Y.; Zhang, Y. C.; Qian, K.; Shen, L.; Phillips, M. I. Protection from ischemic heart injury by a vigilant heme oxygenase-1 plasmid system. *Hypertension* **43**:746–751; 2004.
- [221] Miller, N.; Vile, R. Targeted vectors for gene therapy. *FASEB* **9**:190–199; 1995.
- [222] Kim, S.; Lin, H.; Barr, E.; Chu, L.; Leiden, J. M.; Parmacek, M. S. Transcriptional targeting of replication-defective adenovirus transgene expression to smooth muscle cells in vivo. *J. Clin. Invest.* **100**:1006–1014; 1997.
- [223] Sepulveda, A. R.; Huang, S. L.; Lebovitz, R. M.; Lieberman, M. W. A 346-base pair region of the mouse  $\gamma$ -glutamyl transpeptidase type II promoter contains sufficient cis-acting elements for kidney-restricted expression in transgenic mice. *J. Biol. Chem.* **272**:11959–11967; 1997.
- [224] Ding, Y.; Davison, R. L.; Hardy, D. O.; Zhu, L.-J.; Merrill, D. C.; Catterall, J. F.; Sigmund, C. D. The kidney androgen-regulated protein promoter confers renal proximal tubule cell-specific and highly androgen-responsive expression on the human angiotensinogen gene in transgenic mice. *J. Biol. Chem.* **272**:28142–28148; 1997.
- [225] Korhonen, J.; Lahtinen, I.; Halmekyto, M.; Alhonen, L.; Janne, J.; Dumont, D.; Alitalo, K. Endothelial-specific gene expression directed by the tie gene promoter in vivo. *Blood* **86**:1828–1835; 1995.
- [226] Igarashi, P.; Whyte, D. A.; Li, K.; Nagami, G. T. Cloning and kidney cell-specific activity of the promoter of the murine renal Na-K-Cl cotransporter gene. *J. Biol. Chem.* **271**:9666–9674; 1997.
- [227] Taille, C.; El Benna, J.; Lanone, S.; Dang, M. C.; Ogier-Denis, E.; Aubier, M.; Boczkowski, J. Induction of heme oxygenase-1 inhibits NAD(P)H oxidase activity by down-regulating cytochrome b558 expression via the reduction of heme availability. *J. Biol. Chem.* **279**:28681–28688; 2004.