



Nursing Service Guidelines Endoscopy Unit

Title: **ENDOSCOPE SURVEILLANCE WITH CHANNEL CHECK™**

Responsibility: Endoscopy Staff (RN, LPN & Endo Techs)

Purpose of Guidelines: For detection of three common residual organic soils (blood, protein and carbohydrates) inside endoscopy scopes to help ensure proper cleaning has taken place and to reduce risk of contamination to personnel or patients.

Supplies needed:

1. Channel Check™ testing strips
2. slip tip syringe > 40ml
3. Zip lock baggies
4. 40ml's of sterile water

Procedure: Regular testing of endoscopes will be performed after manual cleaning has taken place in accordance with the manufacturer's guidelines to ensure that the cleaning process is being conducted properly. This is done after manual cleaning, but before High Level Disinfection takes place. Duodenoscopes, all endoscopic ultrasound (EUS) scopes, and all bronchoscopes will be tested after every use. All other scopes will be placed on a monthly rotation check list. Results for all testing will be recorded on the Channel Check™ log sheet.

1. Using a new sterile slip tip syringe, draw 40 ml of commercially available pre-packaged sterile water (i.e., sterile water for irrigation) and 10 ml of air into the syringe. With the distal end of the scope held lower than the biopsy/suction channel, flush with 30 ml of sterile water to rinse out detergent, then place the distal end of the flexible endoscope inside a clean collection container (e.g., sterile urine specimen container or supplied Zip-lock bag).
2. Inject the remaining 10 ml of sterile water through the biopsy/suction channel followed by the 10 ml of air in order to aid in complete flushing of the fluid. Collect all the fluid that drains from the distal end into the sample collection container.
3. Mix the sample well (swishing).
4. With the three pads fully submerged, stir the recovered water for approximately 10 seconds with the test strip.
5. Remove the test strip.
6. Dab the side of the test strip on an absorbent surface (e.g., paper towel) to absorb excess moisture.

7. After 90 seconds, compare the colors on the test strip to the color indicator chart provided.
8. Record the results for all 3 pads. Should any of the pads indicate there is residual soil, re-clean the device and then retest. Record your results in a logbook.
9. Once all three pads are negative, proceed to high level disinfection process.

Control Test: The first step when opening a new bottle of Channel Check™ residual soil test strips is to check the performance of the lot with the included vial of control soil. This will ensure that the reagent in each of the test pads has remained active after shipment. This is only done once per bottle. To test, remove the vial of dehydrated test soil from the box. The test vial holds enough lyophilized test soil to create a single milliliter of test soil.

1. After opening the test strip bottle place a discard date of 90 days from date opened on the sticker provided on the cap.
2. To re-hydrate soil, unscrew the cap from the vial, add exactly 1ml of sterile water to the vial. Screw the cap back on the vial, being sure you have a tight seal.
3. Shake the vial vigorously for at least 1 minute, until a color change is visible in the vial. Check the vial to make sure the soil has been completely re-hydrated.
4. Retrieve a single Channel Check test strip from the pack.
5. Dip the test into the vial, making sure to completely immerse all three test pads into the solution.
6. Swish the test strip in the vial for 10 seconds.
7. After 10 seconds, remove the test strip and dab the side of the moistened test pad on a clean, dry absorbent pad, to dab off excess water.
8. The reagents in the test pads require time to interact with the residual soil, so wait a complete 5 minutes before reading the results.
9. After 5 minutes compare the results to the Control Result Color Chart. The colors of each test pad should closely approximate the colors found on the Control Color Chart.
10. Record the results of the control test on the log sheet.

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